



Les journées du Centre Blaise Pascal  
d'analyse et la modélisation de données issues du monde vivant

# *NMR experiment-driven modeling of biological macromolecules*



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Institut des Sciences Analytiques  
Université de Lyon / UMR 5280 CNRS / ENS Lyon / UCB Lyon 1



$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1-3\cos^2\theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt} \sigma = -i[\mathcal{H}, \sigma]$$

# Outline

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- **General introduction & overview**
  - Experimental techniques for Structural Biology
  - Strength & weakness, complementarity
  - Experimental data content & modeling
- **State-of-the-art liquid and solid-state NMR structure determination**
  - Multi-purpose UNIO platform
  - Recent progress in liquid-state NMR
  - Recent progress in solid-state NMR

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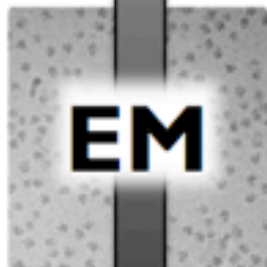
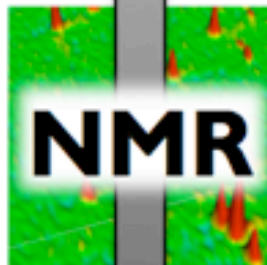
# Experimental techniques & atomic resolution

DTSGTVCL  
SALPPEAT  
DTLNLIAS

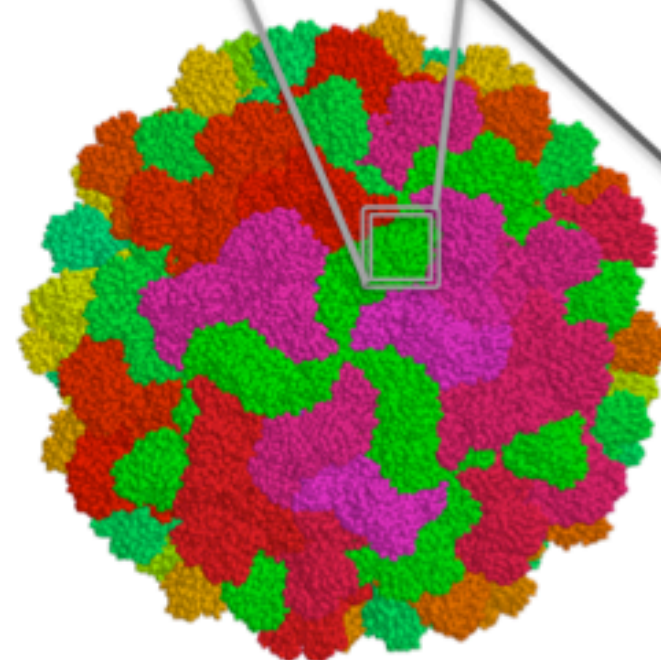
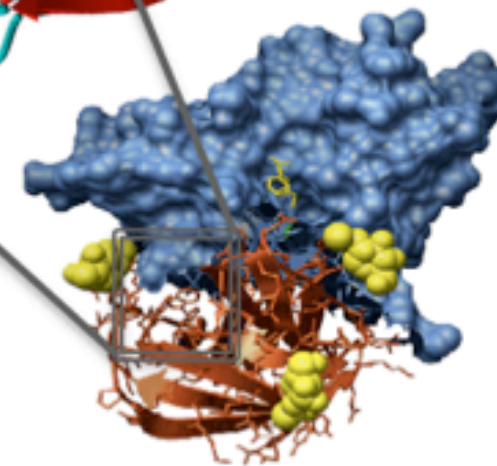
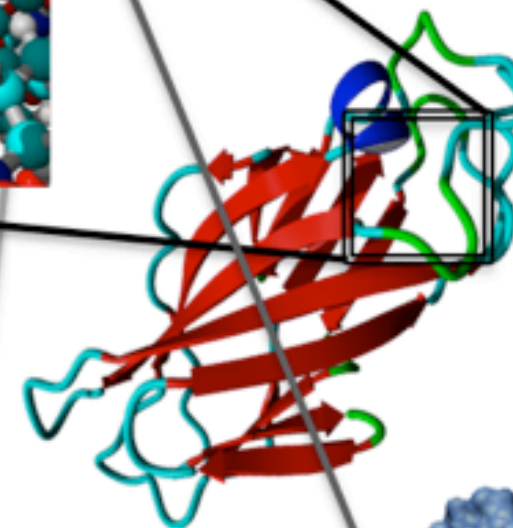
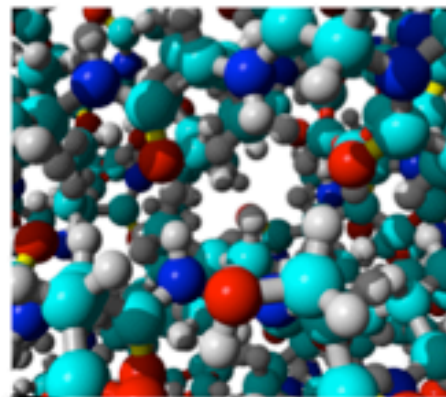


GEATQEDY  
YTGDHYAT  
FSLIDQTC

Higher resolution



ATOM	1	N	ASP	A	1	-10.341	-9.922	9.398
<b>TRANSFORMATION</b>								
ATOM	3	C	ASP	A	1	-11.156	-9.786	8.164
ATOM						-10.288	-9.894	6.915



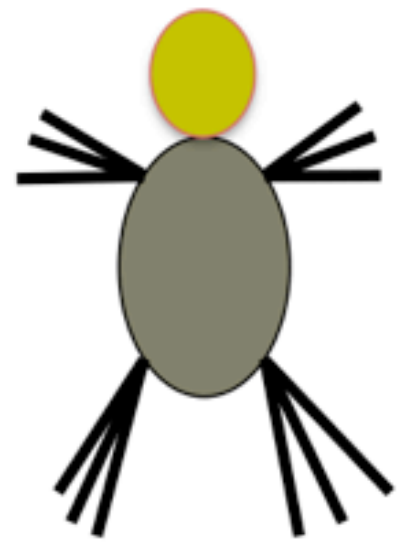
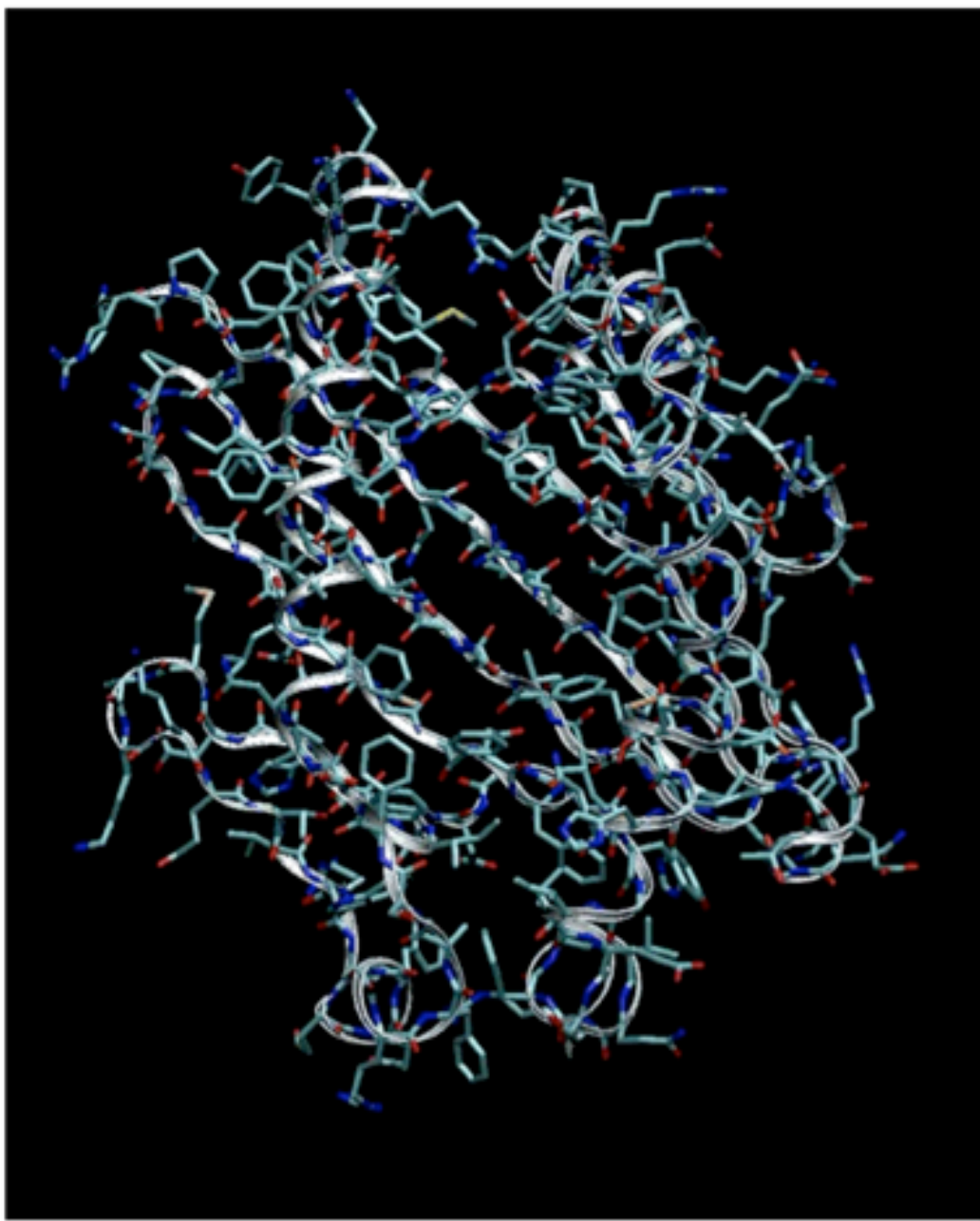
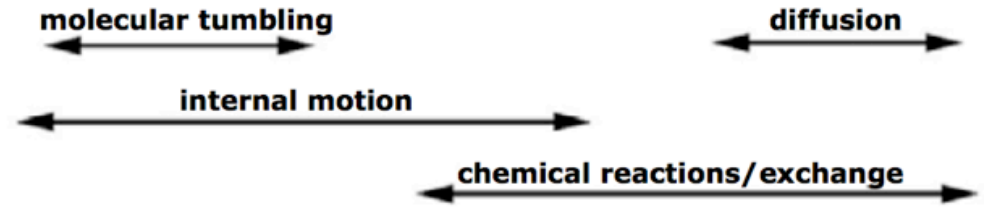
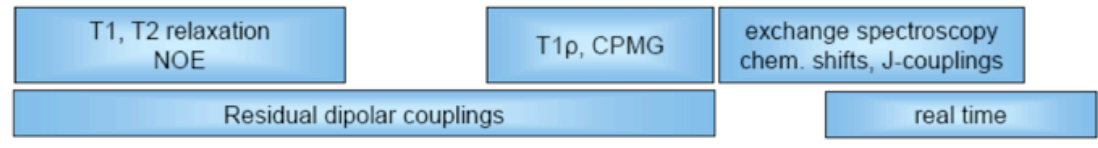
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$$\frac{d}{dt} \sigma = -i[\mathcal{H}, \sigma]$$

*but there is dynamics...*

### Protein dynamics probed by NMR



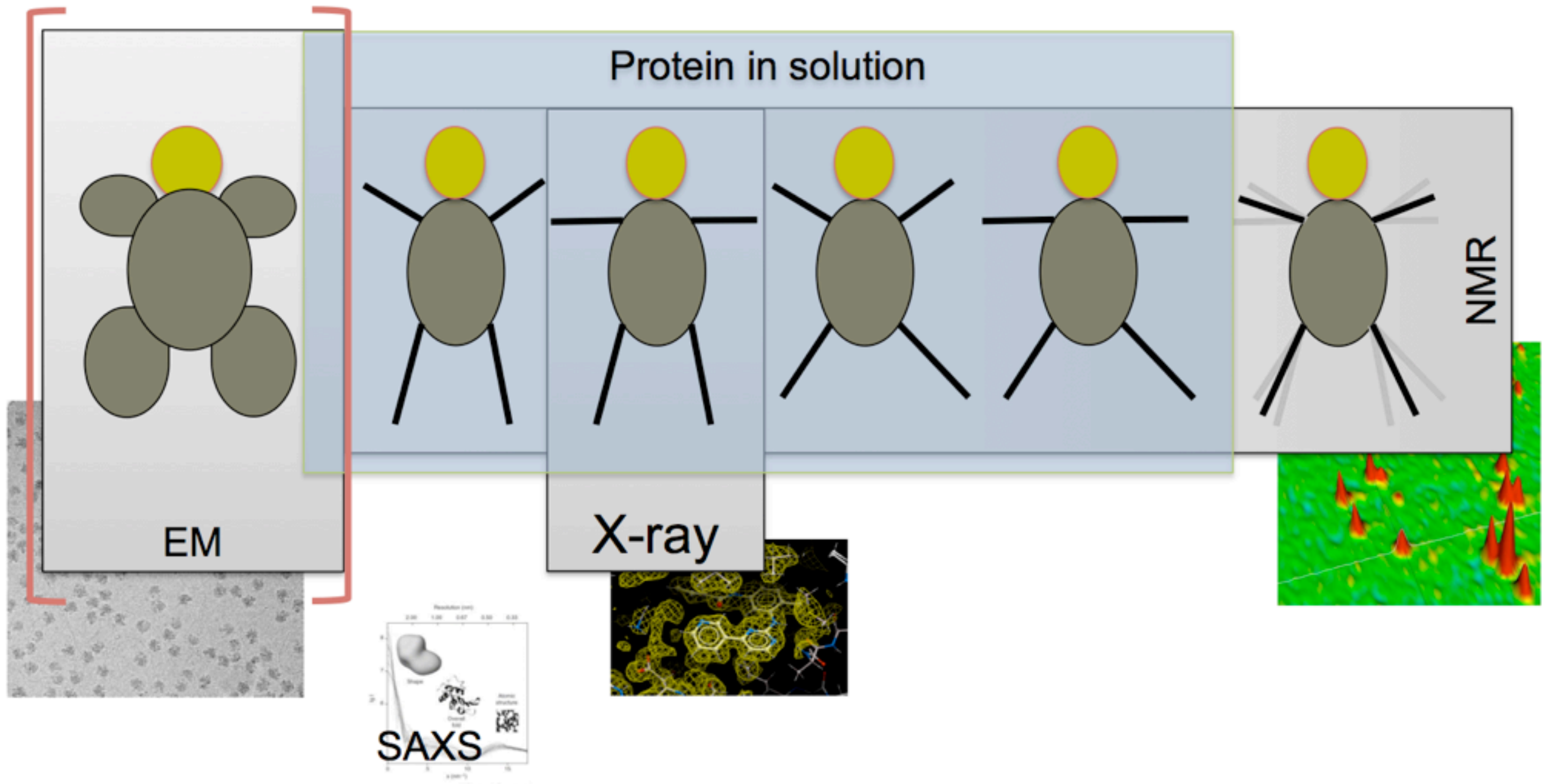
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# Experimental techniques & dynamics

$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\frac{d\sigma}{d\Omega} = -i[\mathcal{H}, \sigma] \mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3\cos^2\theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$



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# What is a macromolecular structure?

$$\frac{d}{dt} \sigma = -i[\mathcal{H}, \sigma]$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

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$$\frac{d}{dt} \sigma = -i[\mathcal{H}, \sigma]$$

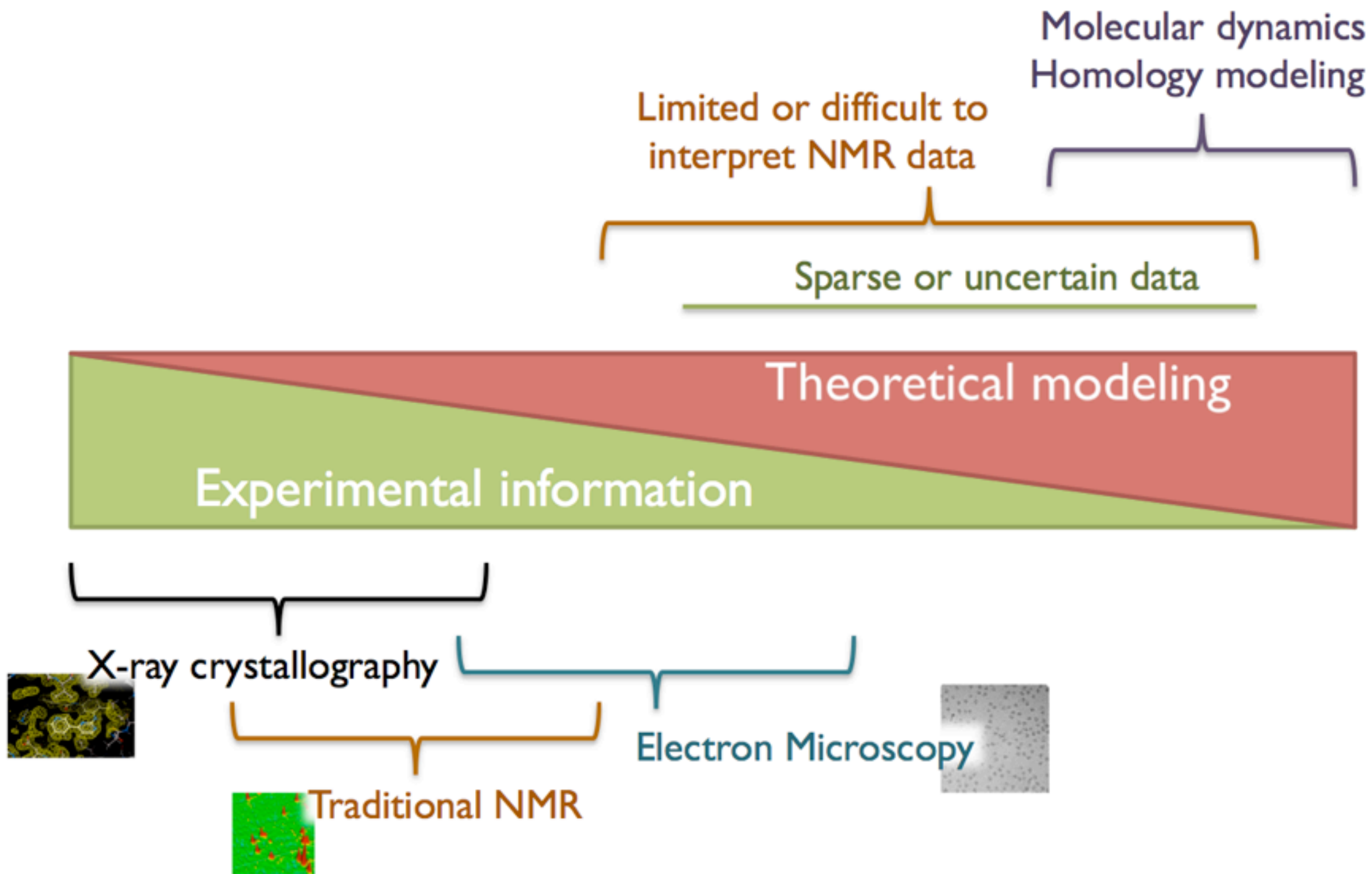
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# History of NMR spectroscopy

## Nobel prizes

1944 *Physics* Rabi (Columbia)



"for his resonance method for recording the magnetic properties of atomic nuclei"

1952 *Physics* Bloch (Stanford), Purcell (Harvard)



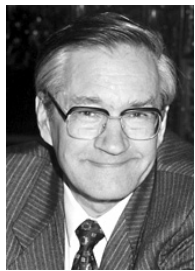
"for their development of new methods for nuclear magnetic precision measurements and discoveries in connection therewith"

2002 *Chemistry* Wüthrich (ETH)



"for his development of nuclear magnetic resonance spectroscopy for determining the three-dimensional structure of biological macromolecules in solution"

1991 *Chemistry* Ernst (ETH)



"for his contributions to the development of the methodology of high resolution nuclear magnetic resonance (NMR) spectroscopy"

2003 *Medicine* Lauterbur (University of Illinois in Urbana ), Mansfield (University of Nottingham)



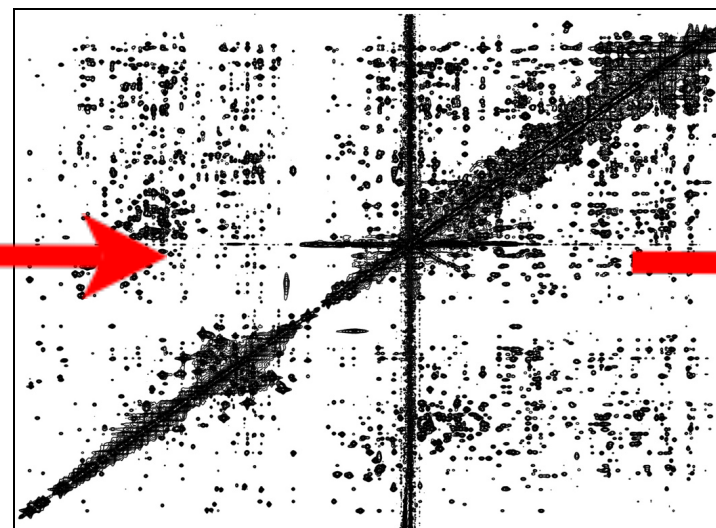
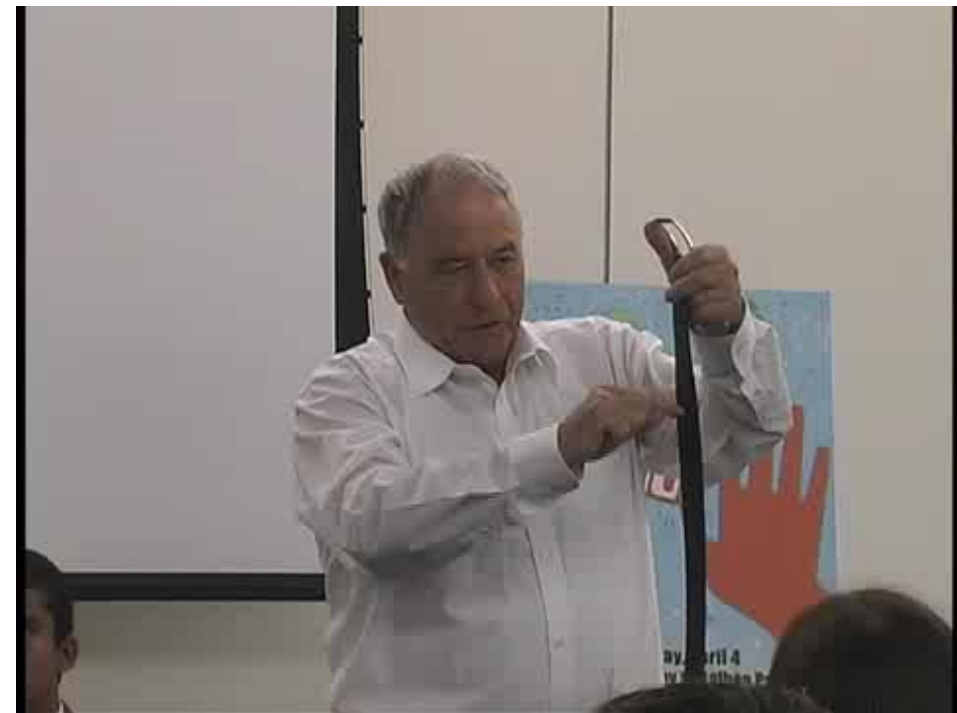
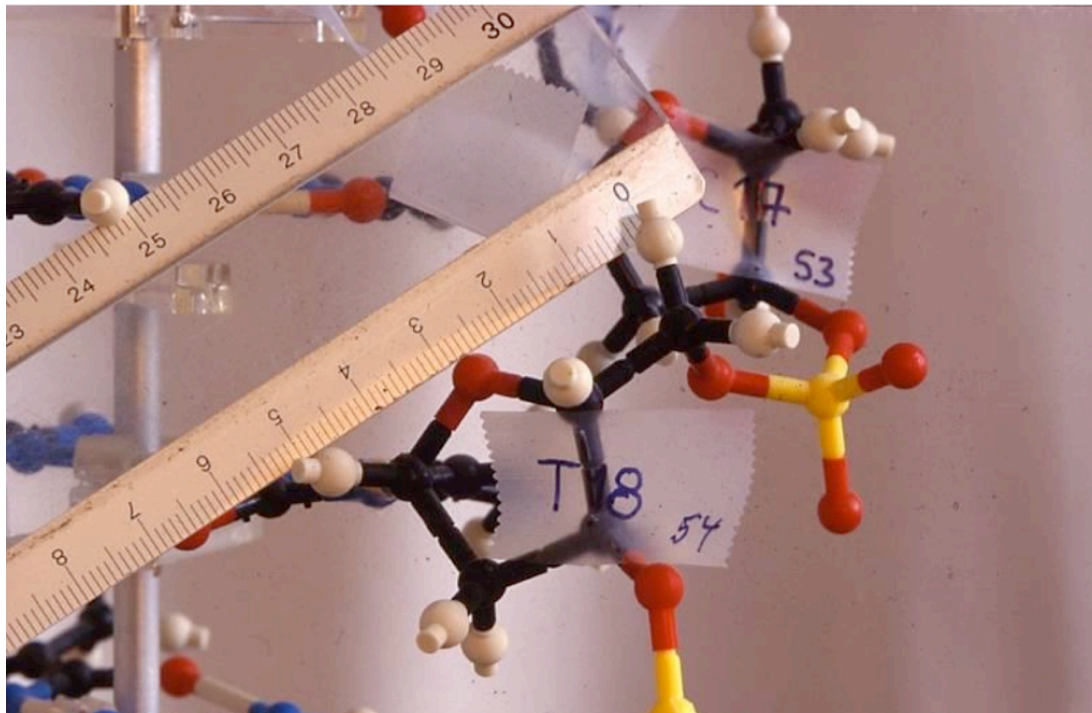
"for their discoveries concerning magnetic resonance imaging"

$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

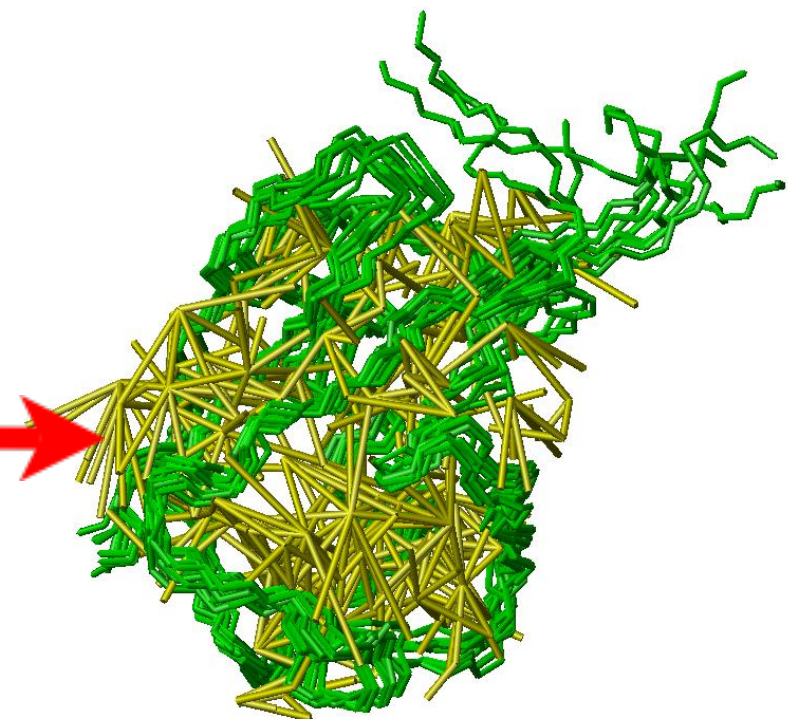
$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

# The basic idea of protein NMR structure determination

It's all about distance measurements between atoms..

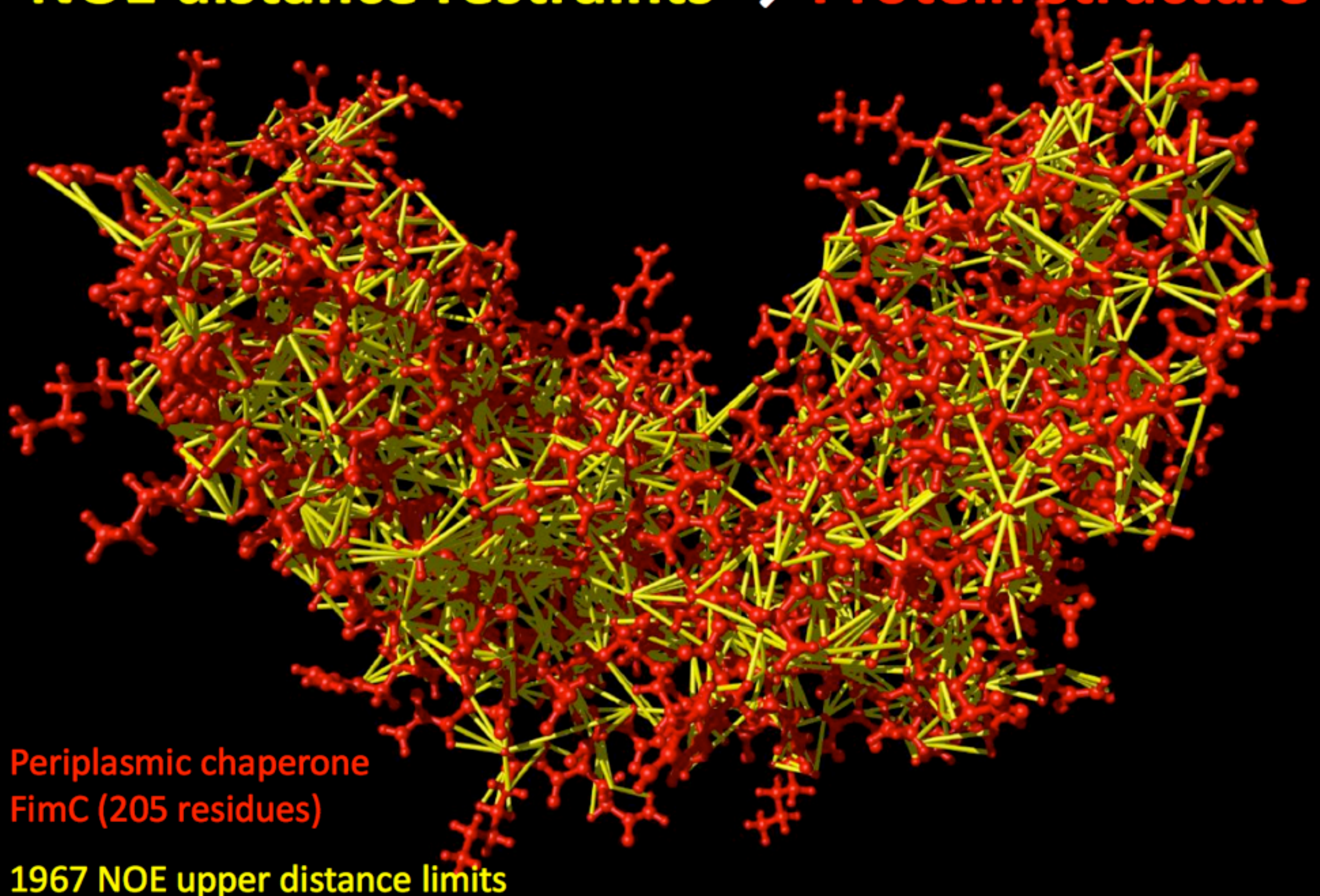


NOESY





# NOE distance restraints → Protein structure



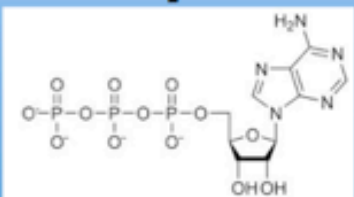
$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

# NMR experiment-driven modeling of macromolecules

DTSGTVCLSALPPEATDTLNLIASDGPFPYSQ  
 DGVVFQNRRESVLPQTQSYGYHYEYTVITPGART  
 RGTRRIITGEATQEDYYTGDHYATFSLIDQTC

## Description of molecules



Translate

Force field (physical)  
 Force field (experimental)

```

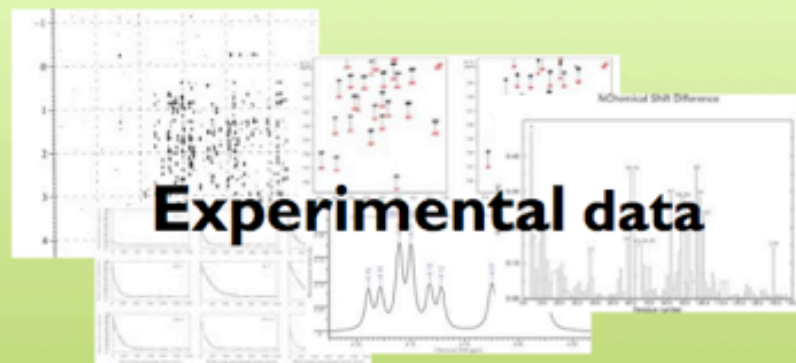
if ($whichMol == "Cartesian") then
  flags excl
  dihed end
  ##SCRIPTS:
  #ocols/sa_l_reduced.cns { defines storel }
  #SCRIPTS:
  #ocols/sa_l_hightemp.cns(nstep=$high_steps;SaProtocol=$SaProtocol; Data=$Data;

else !torsion
  single dynamics
  ##SCRIPTS:
  #ocols/torsiontop.cns

(* 1 ===== high temperature dynamics *)
parameter
  b
  (name sg) (name sg) 0.0 TOKEN
  e
  (all) (name sg) (name sg) 0.0 TOKEN
  b
  (name C1') (name C2') 5.0 TOKEN
end
##SCRIPTS:
#ocols/sa_l_reduced.cns { defines storel }
flags exc
dihed end
##SCRIPTS:
#ocols/sa_l_tad_hightemp.cns
(nstep=$high_steps;SaProtocol=$SaProtocol;Data=$Data;)

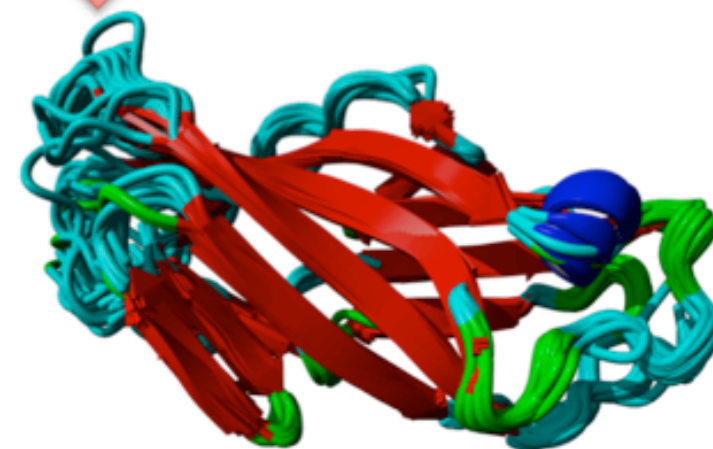
(* 2 ===== cooling 1 *)
##SCRIPTS:
#ocols/sa_l_tad_cool1.cns(SaProtocol=$SaProtocol;Data=$Data;Toppar=$Toppar;)
! Here part
TAD ends
end if
  
```

## Calculation protocol



## Experimental data

Translate



$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

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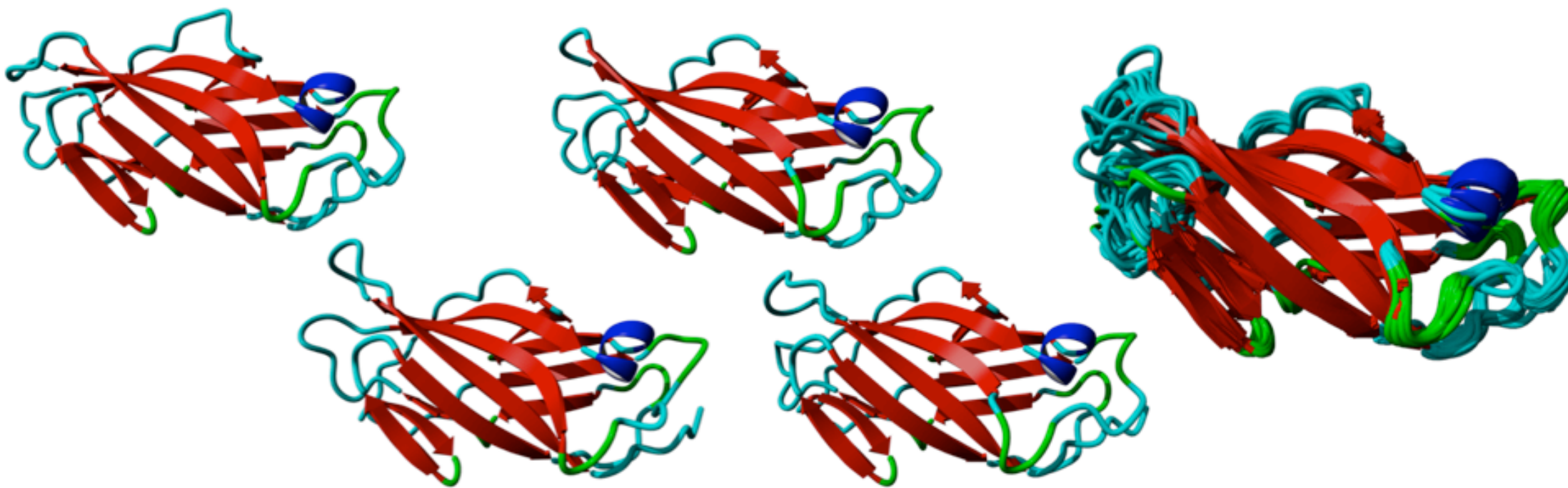
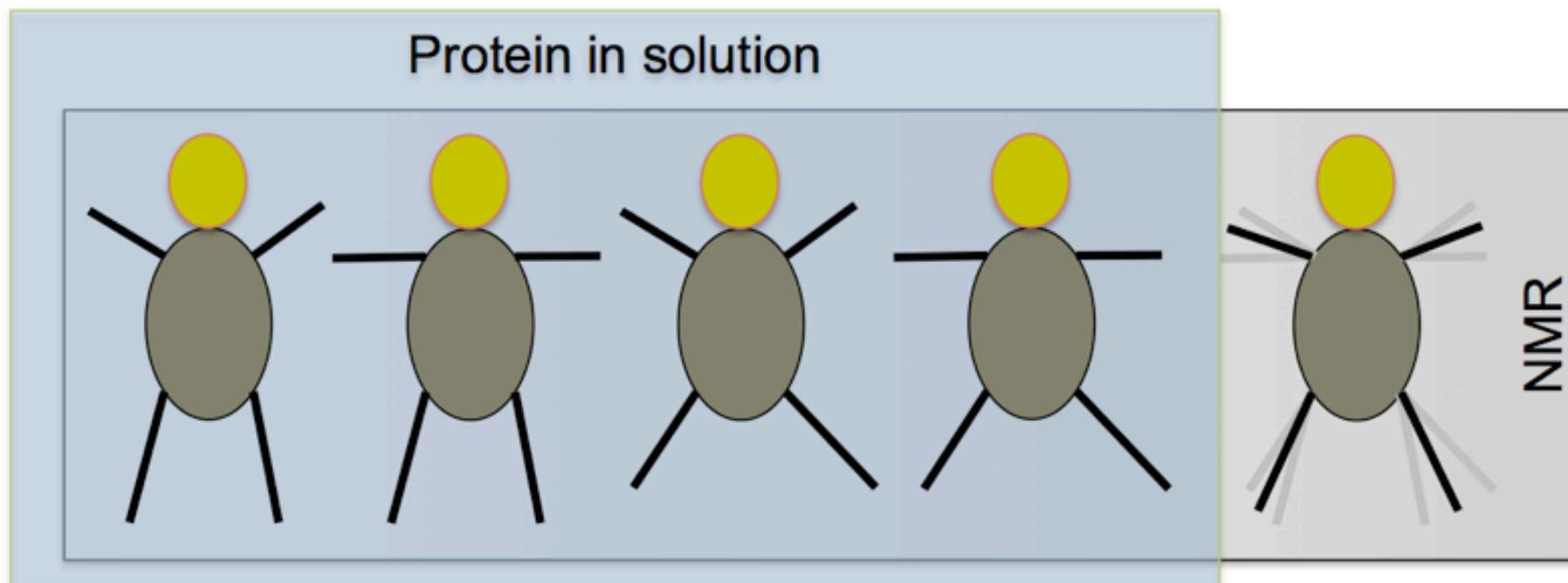
# Averaged NMR data & dynamics

$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1-3\cos^2\theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

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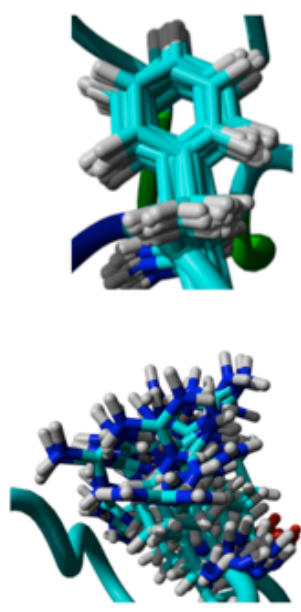
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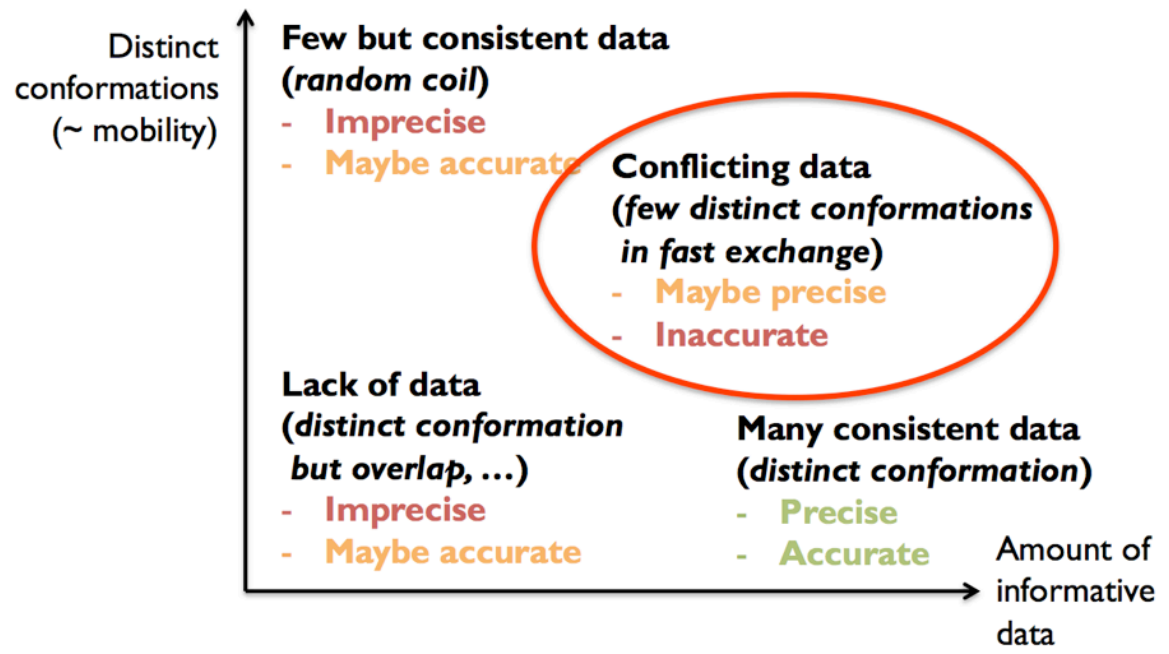
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# Accuracy vs. precision

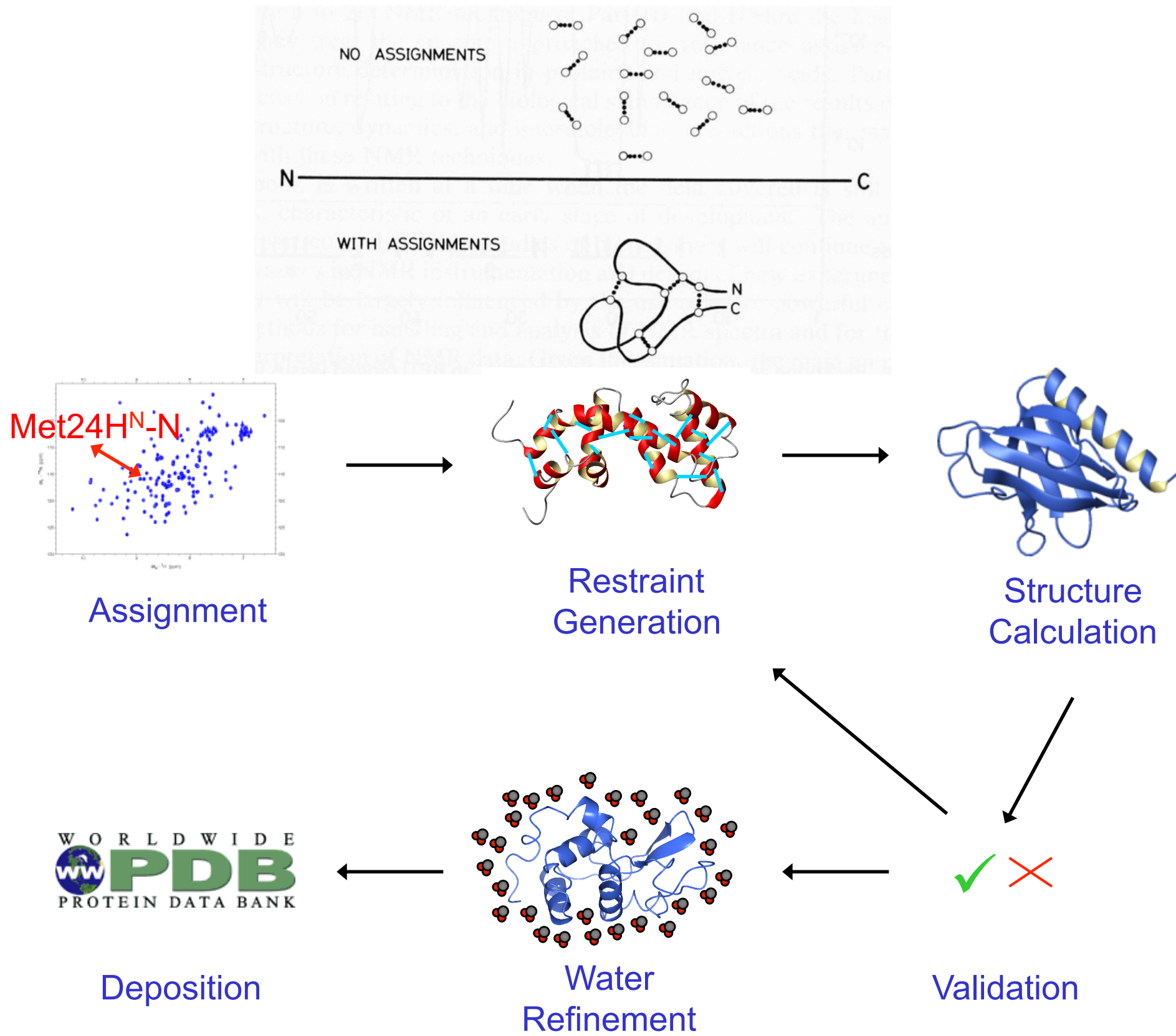


	Accurate	Inaccurate (systematic error)
Precise <b>Low RMSD</b>		
Imprecise (reproducibility error) <b>High RMSD</b>		

## Amount of data available vs. conformations



# Protocol for protein NMR structure determination



$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

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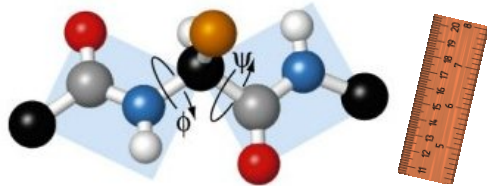
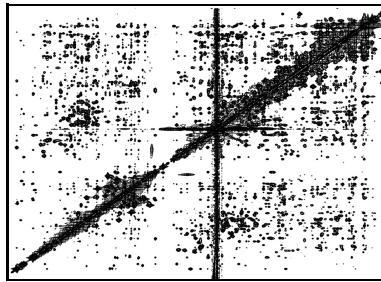
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  - Recent progress in solid-state NMR

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$$(\omega_1, \omega_2) = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

# Research overview: unsupervised molecular modeling



Protein Sequence

...KLTDSQNFDEYMKALGVGFATRQVGNMKTITLNGSQQEGGKVM...

Computational methods



Protein Structure Model



$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_N \hbar^2 \gamma_1 \gamma_2}{\lambda \pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt} \sigma = -i [\mathcal{H}, \sigma]$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_N \hbar^2 \gamma_1 \gamma_2}{\lambda \pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

*What is the challenge?*

**NMR resonance assignment is like solving a puzzle...**

**...with missing pieces  
(incomplete signals)**



**...with additional pieces  
(artifacts)**



**...in the mist  
(low signal-to-noise,  
line-broadening)**



$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

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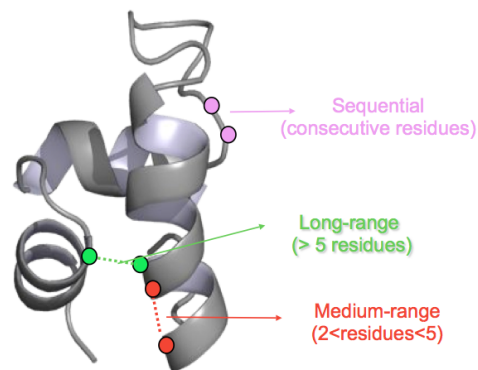
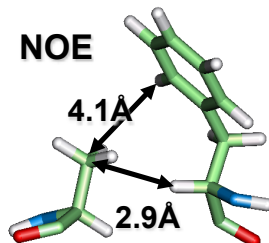
# What is the challenge?

## NMR - Giant Jigsaw Puzzle

### NMR Structure Determination

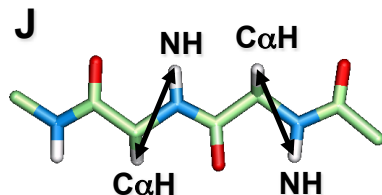
#### NOE

- a through space correlation (<5Å)
- distance constraint



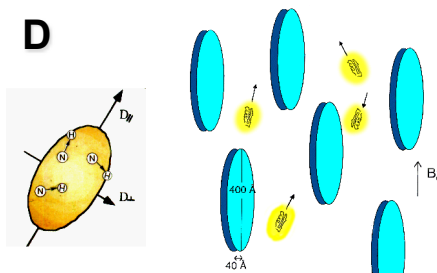
#### Coupling Constant (J)

- through bond correlation
- dihedral angle constraint



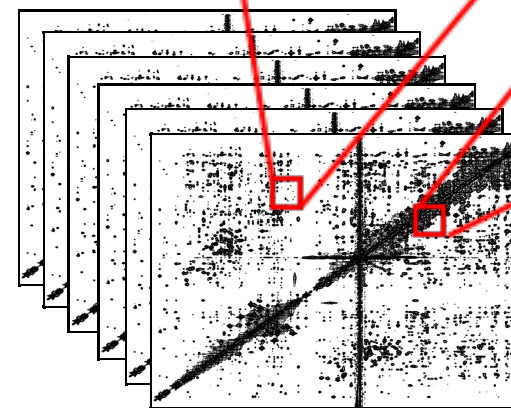
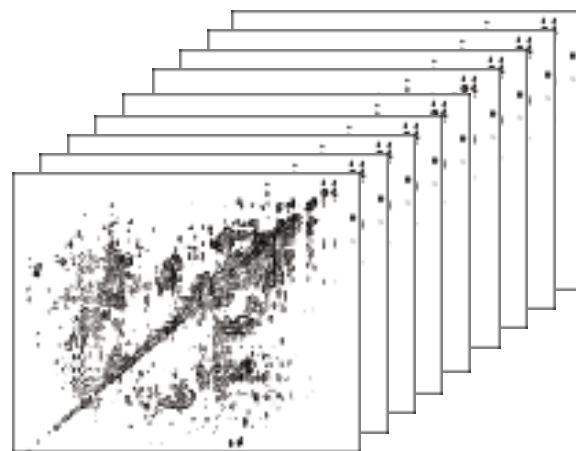
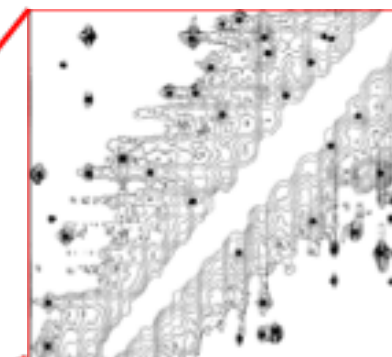
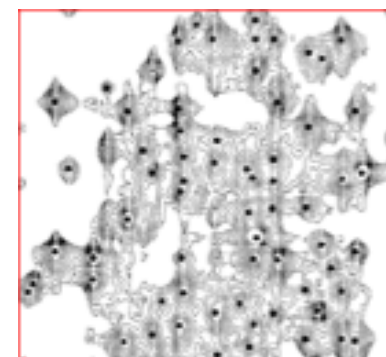
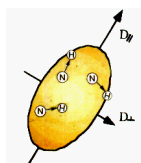
#### Chemical Shift

- very sensitive to local changes in environment
- dihedral angle constraint



#### Dipolar coupling constants (D)

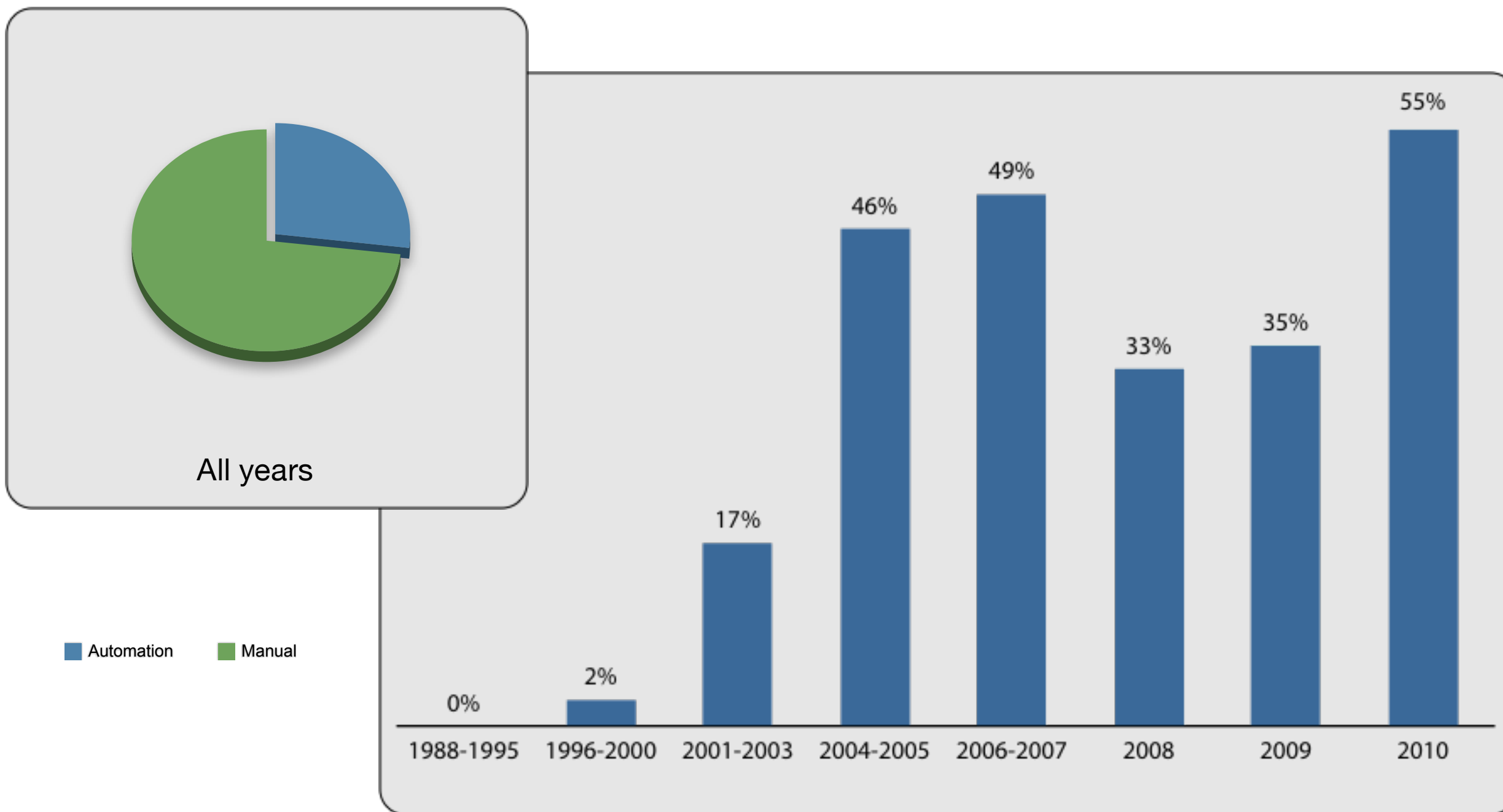
- bond vector orientation relative to magnetic field
- alignment with bicelles or viruses



Often **15-20** NMR spectra need to be consistently and jointly analyzed.

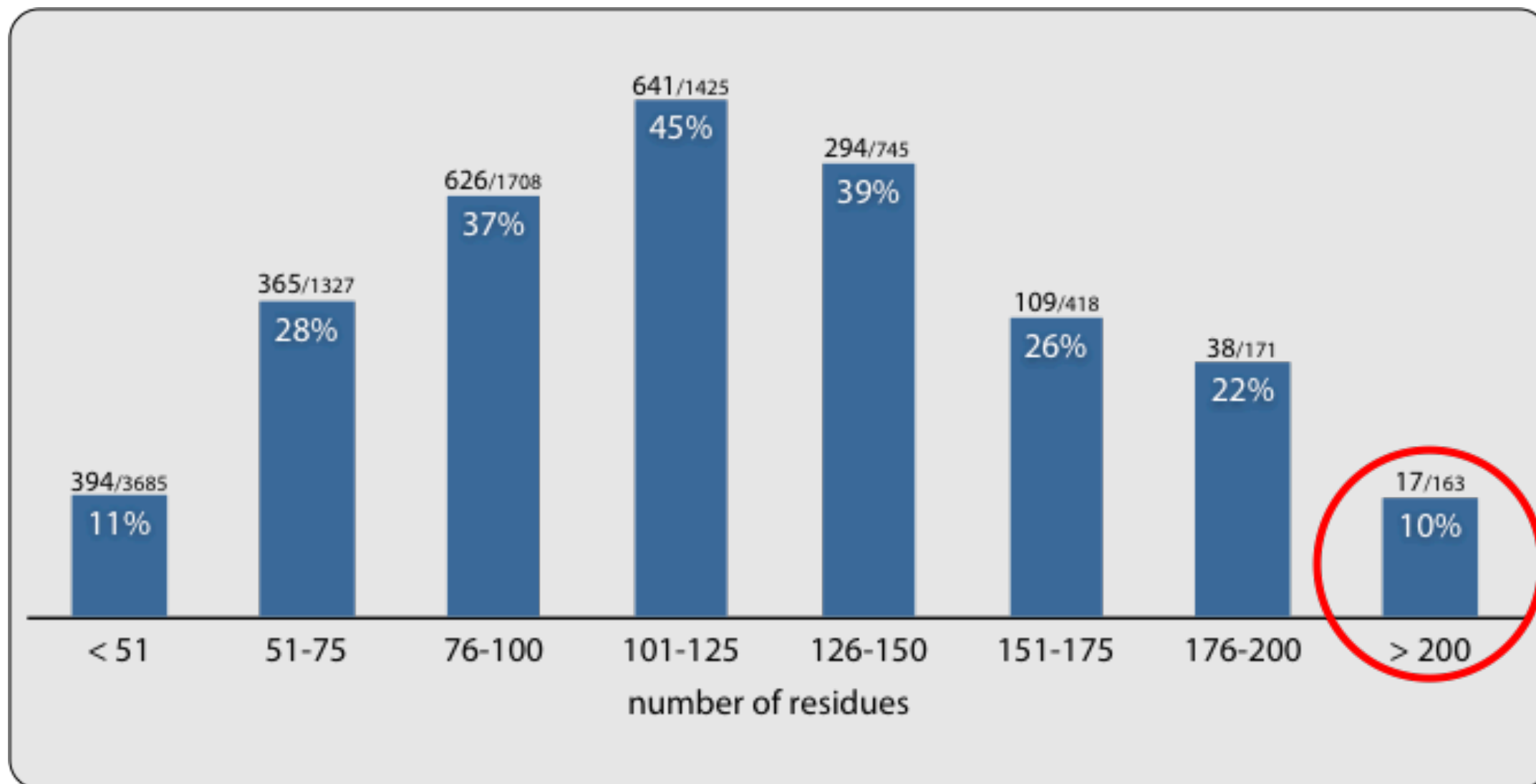
# What is the challenge?

Percentage of “automated” PDB-deposited NMR structures



# What is the challenge?

Size of “automated” PDB-deposited NMR structures



**! Need of improvement for ‘traditional’ algorithms !**

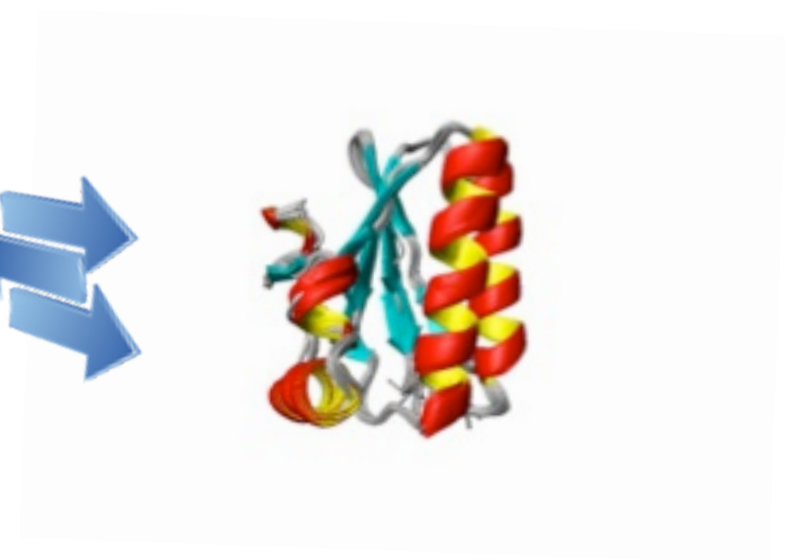
$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} \frac{\hbar^2 \gamma_1 \gamma_2}{3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt} \sigma = -i [\mathcal{H}, \sigma]$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} \frac{\hbar^2 \gamma_1 \gamma_2}{3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

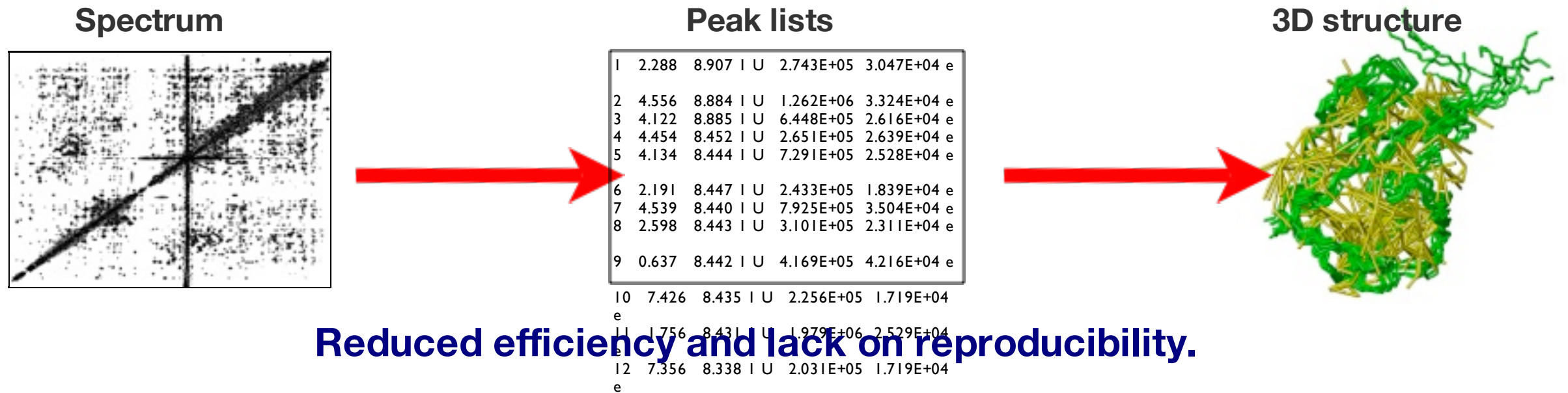
# Outline - Part 1



**UNIO**

# Aim of UNIO: Overcome current weaknesses ...

- Automated approaches operate on intermediate textual peak lists.



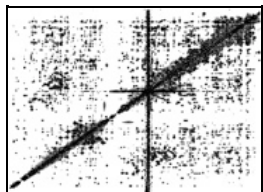
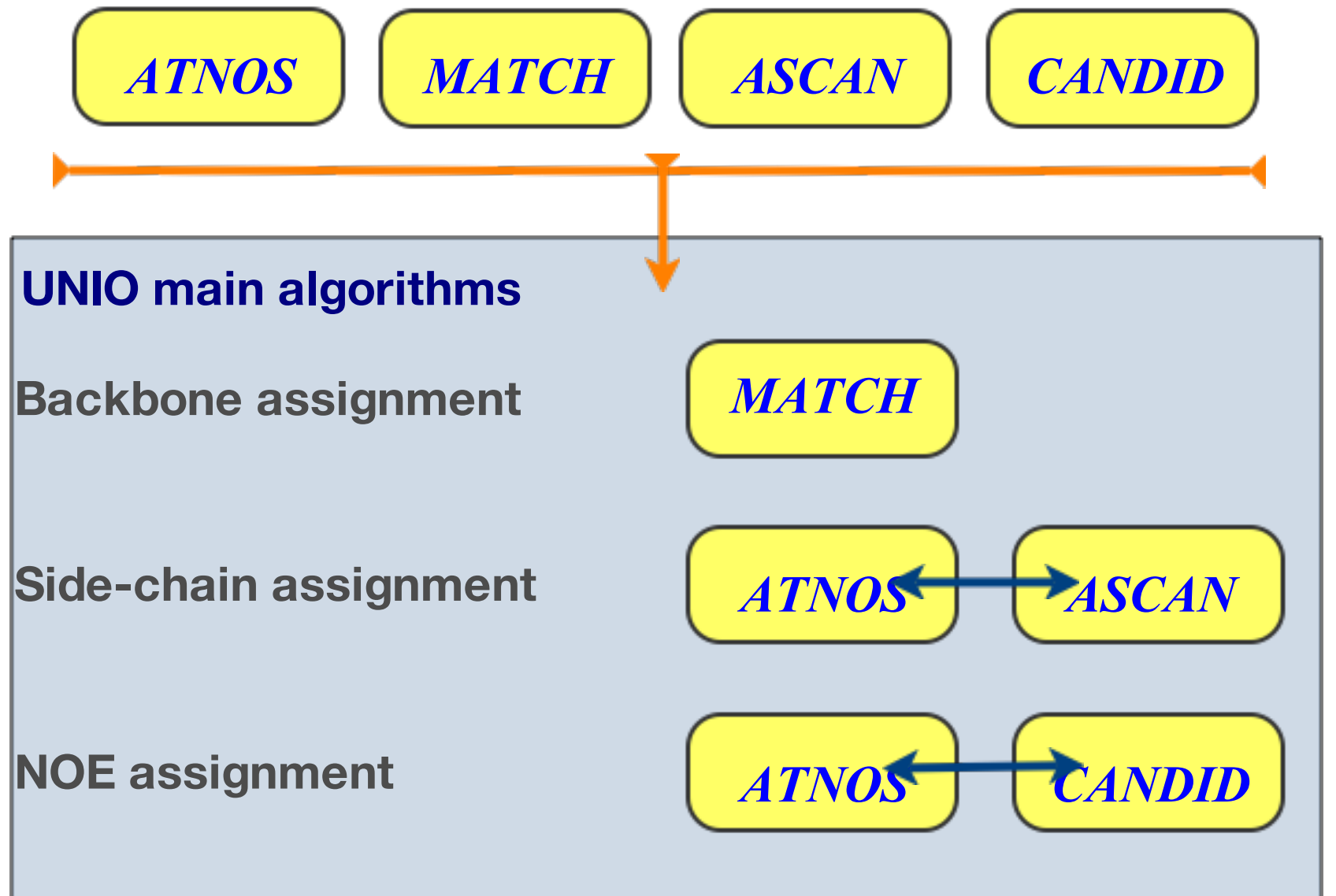
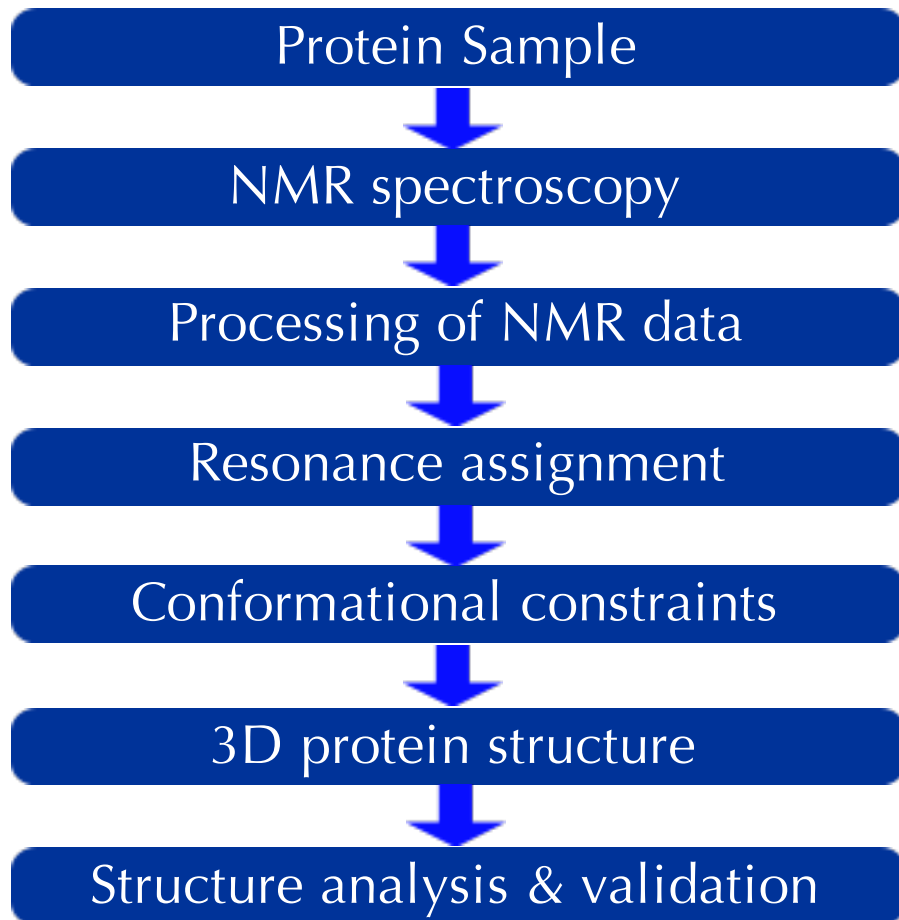
- Automated approaches target individual stages.



NMR is a **multi-stage** data analysis process  
**Brute force automation** is likely to fail.

**Smooth integration** is key for laboratory efficiency.

# UNIO: a multi-purpose application suite

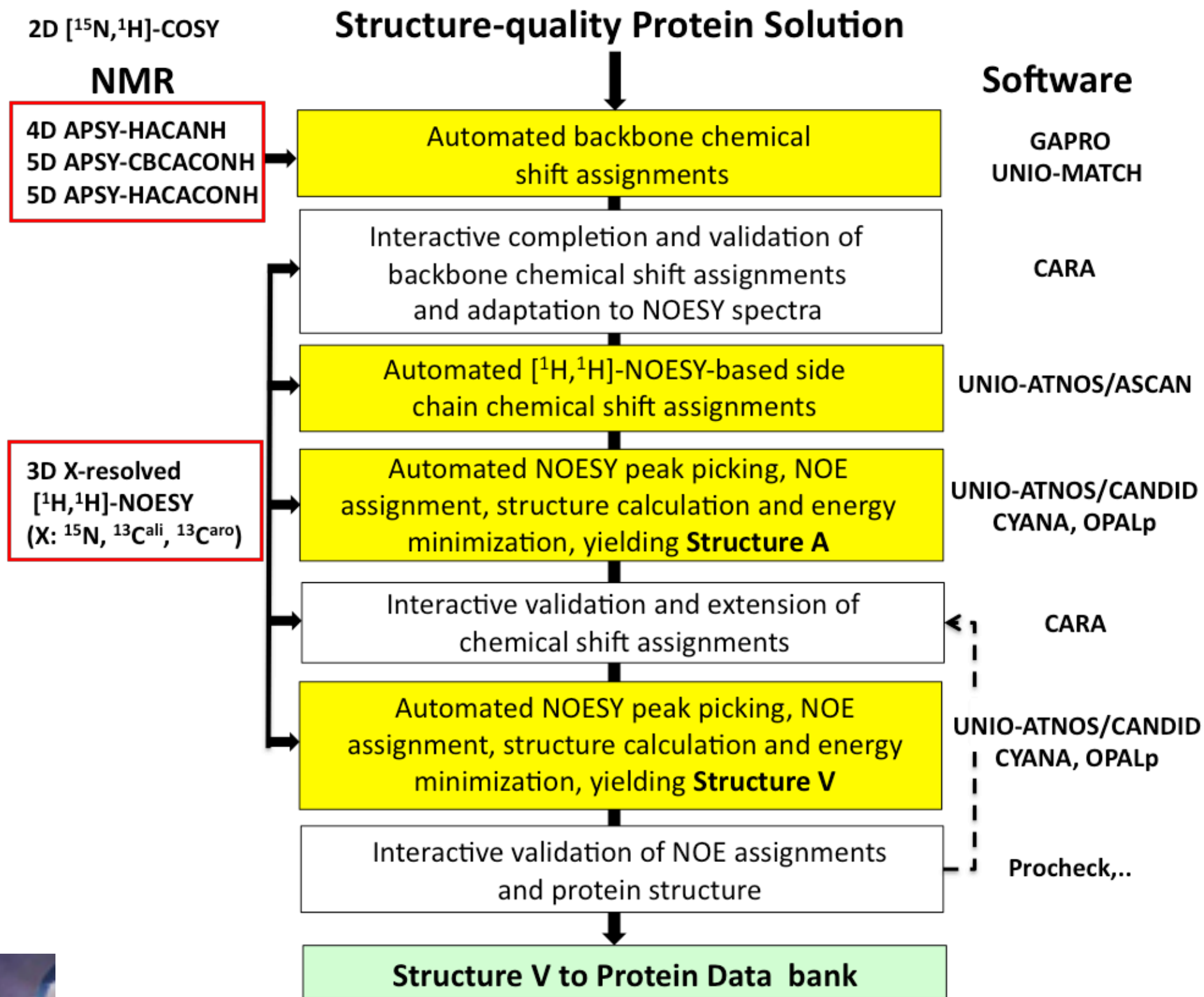


- UNIO assembles four previously developed expert algorithms into three principal data analysis modules, each one **operating directly on the NMR spectra**.



- Correct and consistent chemical shift referencing of all NMR spectra.
- Adaptation of previous chemical shifts to subsequently used set of NMR spectra.

# Application example: The J-UNIO protocol



$$\omega_1, \omega_2) = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$I(\omega_1, \omega_2) = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

# J-UNIO protocol: a representative example

## Initial Phase 1

**TM1367**

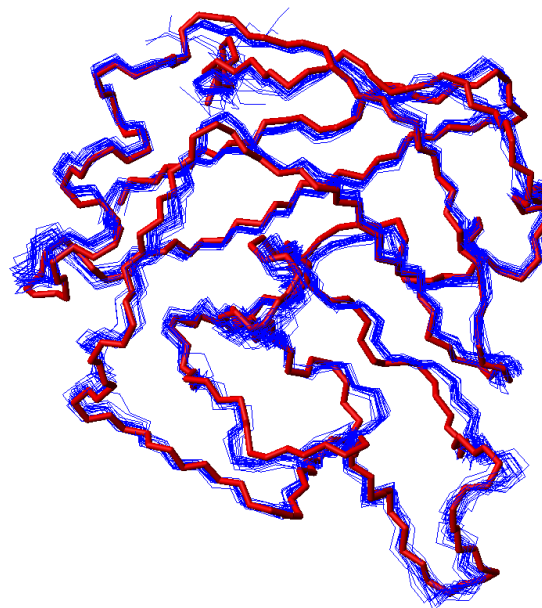
124 aa

NMR data collection and  
automated NMR structure  
determination

7d

RMSD to X-ray structure  
 $1.11 \pm 0.10 \text{ \AA (bb)}$

**Starting Structure A**





# Real-world UNIO application: an example

## Initial Phase 1

**TM1367**  
124 aa

NMR data collection and  
automated NMR structure  
determination

7d

RMSD to X-ray structure  
 $1.11 \pm 0.10 \text{ \AA (bb)}$

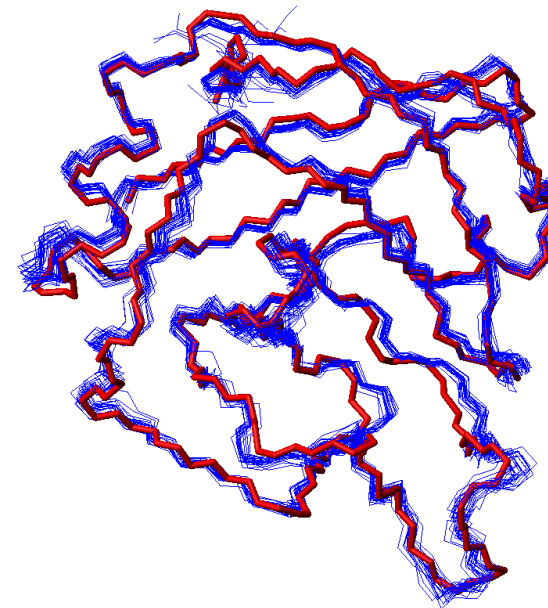
## Refinement Phase 2

Interactive NMR  
structure refinement

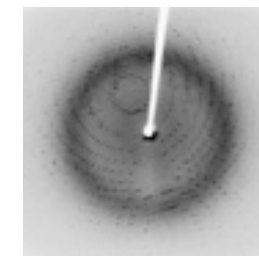
6d

RMSD to X-ray structure  
 $0.92 \pm 0.06 \text{ \AA (bb)}$   
 $1.54 \pm 0.06 \text{ \AA (ha)}$

**Final Structure V**



Similar to the work-flow seen in X-ray



# PDB structures determined by the J-UNIO protocol

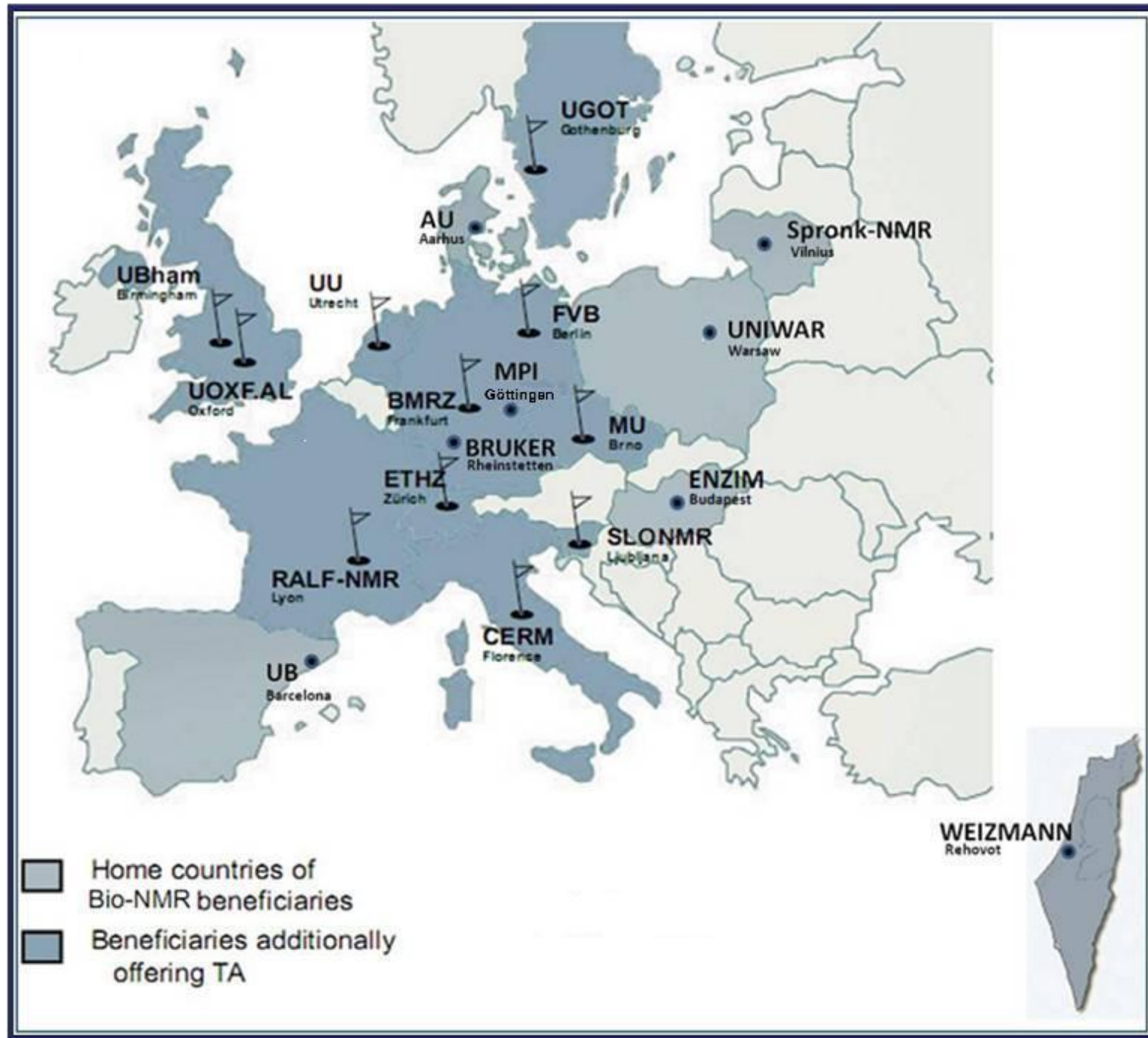
Initial Phase 1 versus Refinement Phase 2

Protein sample (PDB id) <sup>a</sup>	Size (aa)	Chemical shift assignments				Precision (bb RMSD, Å)			RMSD <sub>AV</sub> <sup>i</sup>
		H $\alpha$ ,C $\alpha$ ,H <sup>N</sup> ,N,C',C $\beta$ (%)		All-atoms <sup>b</sup> (%)		Residues <sup>f</sup>	ASCAN <sup>g</sup>	Final <sup>h</sup>	
		MATCH <sup>c</sup>	Final <sup>d</sup>	ASCAN <sup>e</sup>	Final <sup>d</sup>				
<b>17 (37)</b>									
TM1112 (2k9z)	89	96	100	72	90	2-89	0.79	0.43	1.45
TM0212 (2ka7)	124	100	100	66	92	1-110	0.69	0.51	1.16
TM1367 (2ka0)	124	92	98	80	92	2-123	0.80	0.44	2.28
A2LD1 (2kl2)	149	85	97			2-100,106-144			
YP_001336205.1 (2l1s)	83	82	99	89	95	4-82	0.73	0.44	1.23
TM0320 (2kyz)	67	91	97	80	96	1-67	0.57	0.45	0.81
YP_510488.1 (2kzc)	85	93	96	76	94	1-85	1.17	0.68	1.41
NP_415897.1 (2kts)	117	81	100	76	93	3-117	1.65	0.62	1.64
YP_399305.1 (2l1n)	120	82	99	67	94	1-34,43-92,97-117	2.03	0.58	1.75
NP_954075.1 (2l1t)	109	91	98	78	95	7-103	0.80	0.64	1.14
NP_253742.1 (2l6p)	124	81	95	76	91	2-38,49-117	1.08	0.61	1.17
YP_001092504.1 (2l6n)	132	76	96	78	93	8-44, 56-120	2.42	0.70	2.82
YP_926445.1 (2l6o)	114	92	98	67	95	10-40,44-113	1.98	0.64	2.32
NP_888769.1 (2l25)	141	87	99	78	93	3-50, 66-136	1.78	0.71	2.01
YP_546394.1 (2l9d)	108	95	99	75	95	8-108	1.89	0.67	2.01
YP_557733.1 (2la7)	145	75	100	80	96	18-144	1.37	0.58	1.97
YP_001302112.1 (2lg7)	129	92	100	81	93	11-81, 91-129	1.45	0.75	2.20

$$\omega_1, \omega_2) = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

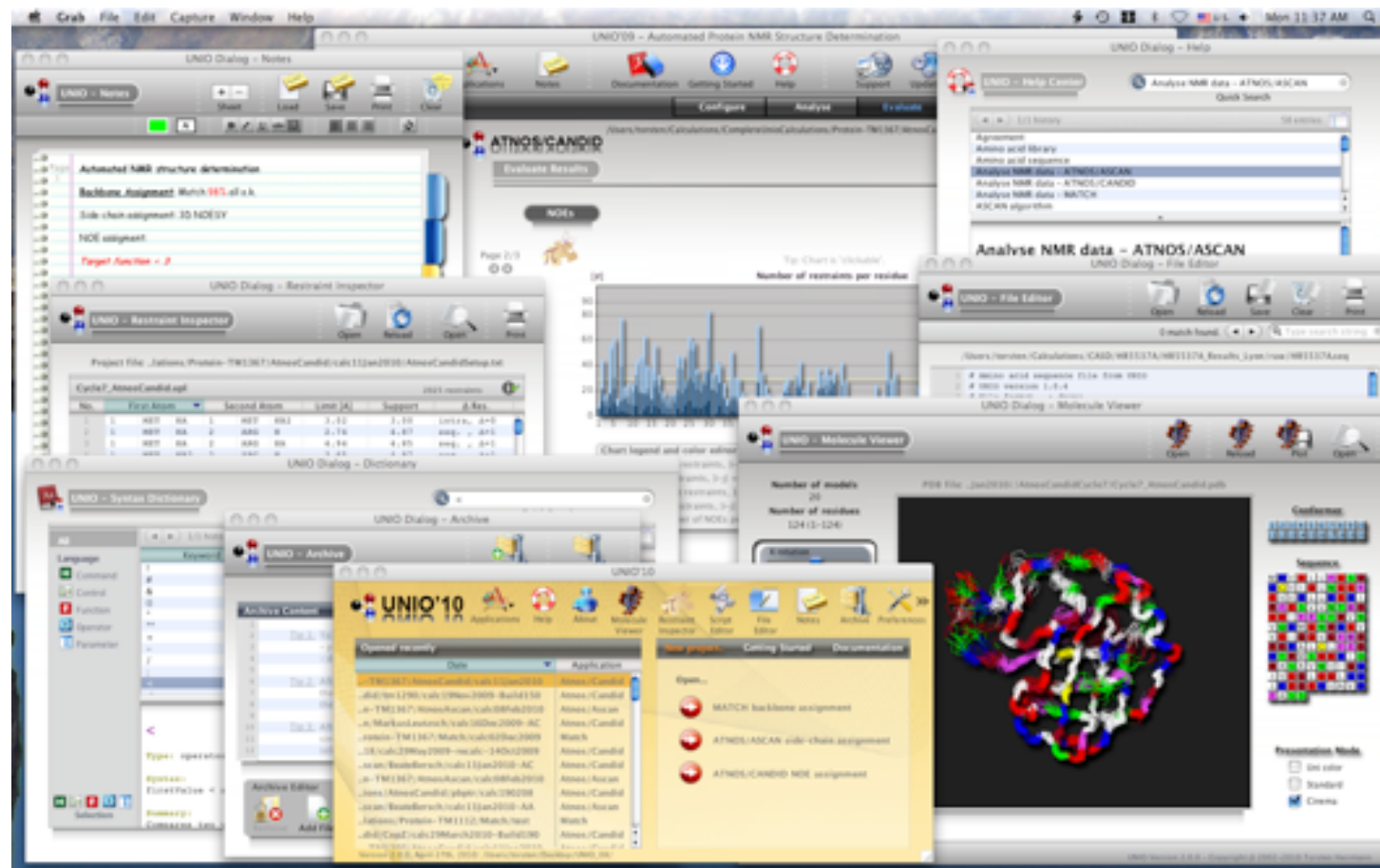
$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

# RALF-NMR offers the J-UNIO protocol



*Financial support by the Access to Research Infrastructures activity in the 7th Framework Programme of the EC (Project number: 261863, Bio-NMR) for conducting the research is gratefully acknowledged*

*But UNIO is **more** - Multipurpose data analysis platform*



One of the major strengths and specificities of UNIO is its **flexibility**. Modules can be launched separately, out of sequence and with different input data. Therefore the UNIO setup can be tailored **for any structure determination project** according to a particular problem at hand.



**Create your own protocol**



$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda} \frac{\hbar^2 \gamma_1 \gamma_2}{\pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt} \sigma = -i [\mathcal{H}, \sigma]$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda} \frac{\hbar^2 \gamma_1 \gamma_2}{\pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

# CASD-NMR

## Critical Assessment of automated Structure Determination of proteins from NMR data

Community-wide initiative for assessing the feasibility of obtaining in an **unsupervised manner** solution NMR structures of proteins with a quality for direct deposition into the Protein Data Bank.

### CASD-NMR 1 (June 2009 - Feb 2010)

10 blind structure determinations of proteins

protein sequence, resonance assignment and **unassigned, refined peak lists**

### CASD-NMR 2 (March 2011-present)

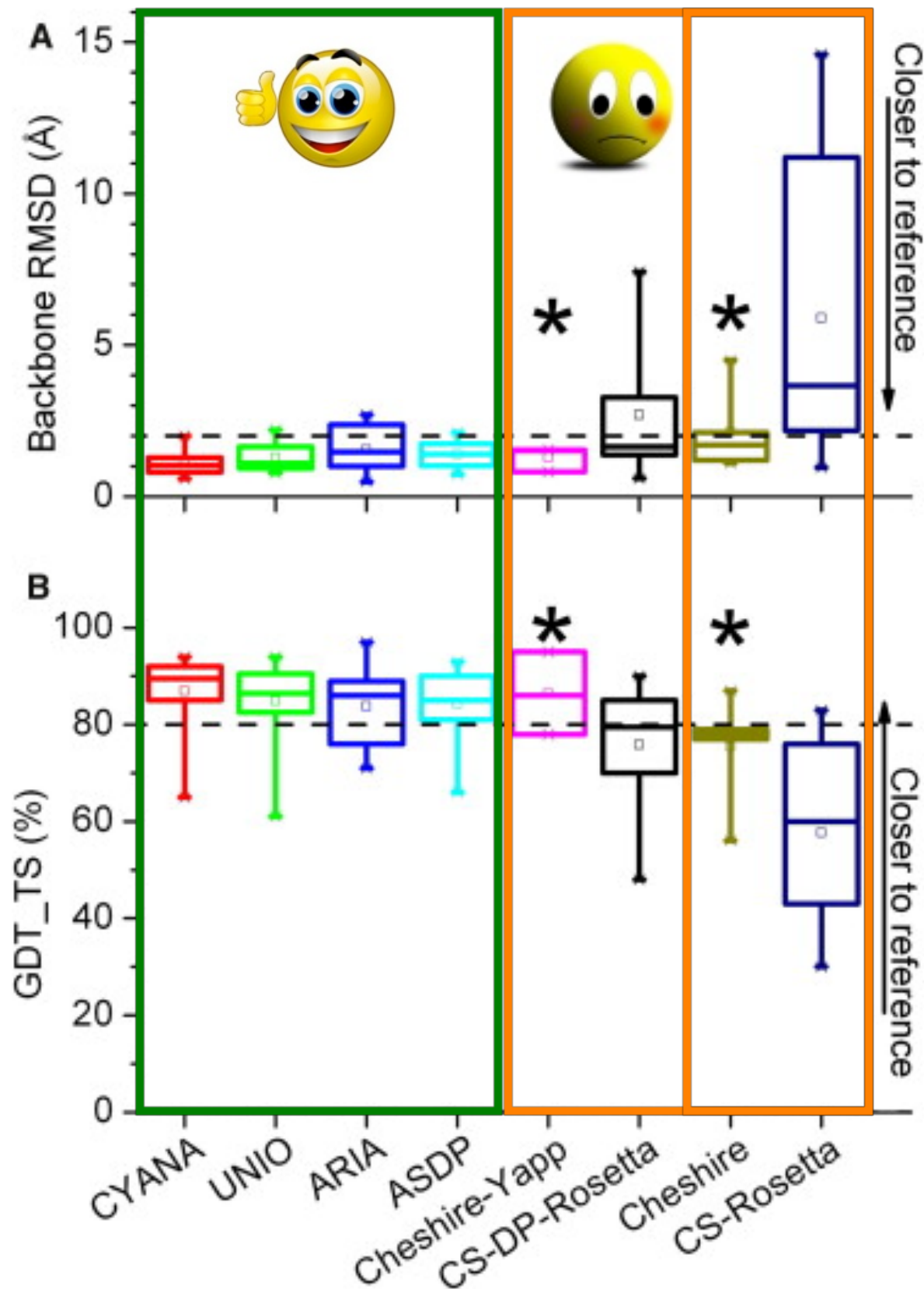
8 blind structure determinations of proteins (so far)

protein sequence, resonance assignment and **NMR spectra**



Deploying & unifying the computational NMR infrastructure

# CASD-NMR I: Results & Conclusions



- Automated NOE assignment yields structures **< 2 Å** from the reference
- Chemical-shift based calculations show high variability in their performance. Filtering with NOESY data might help sometimes.

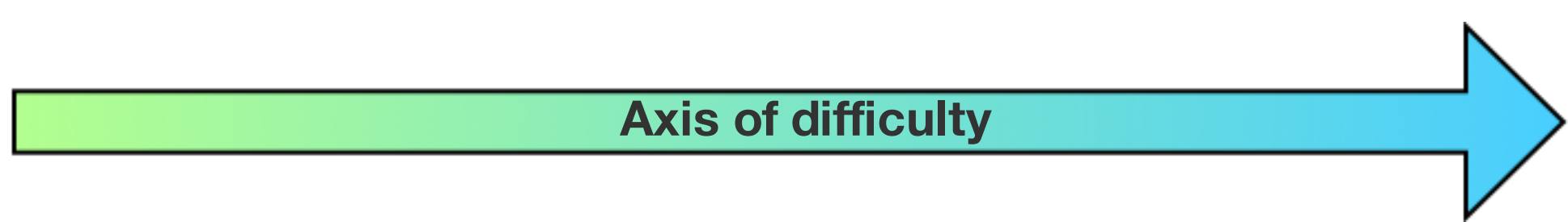
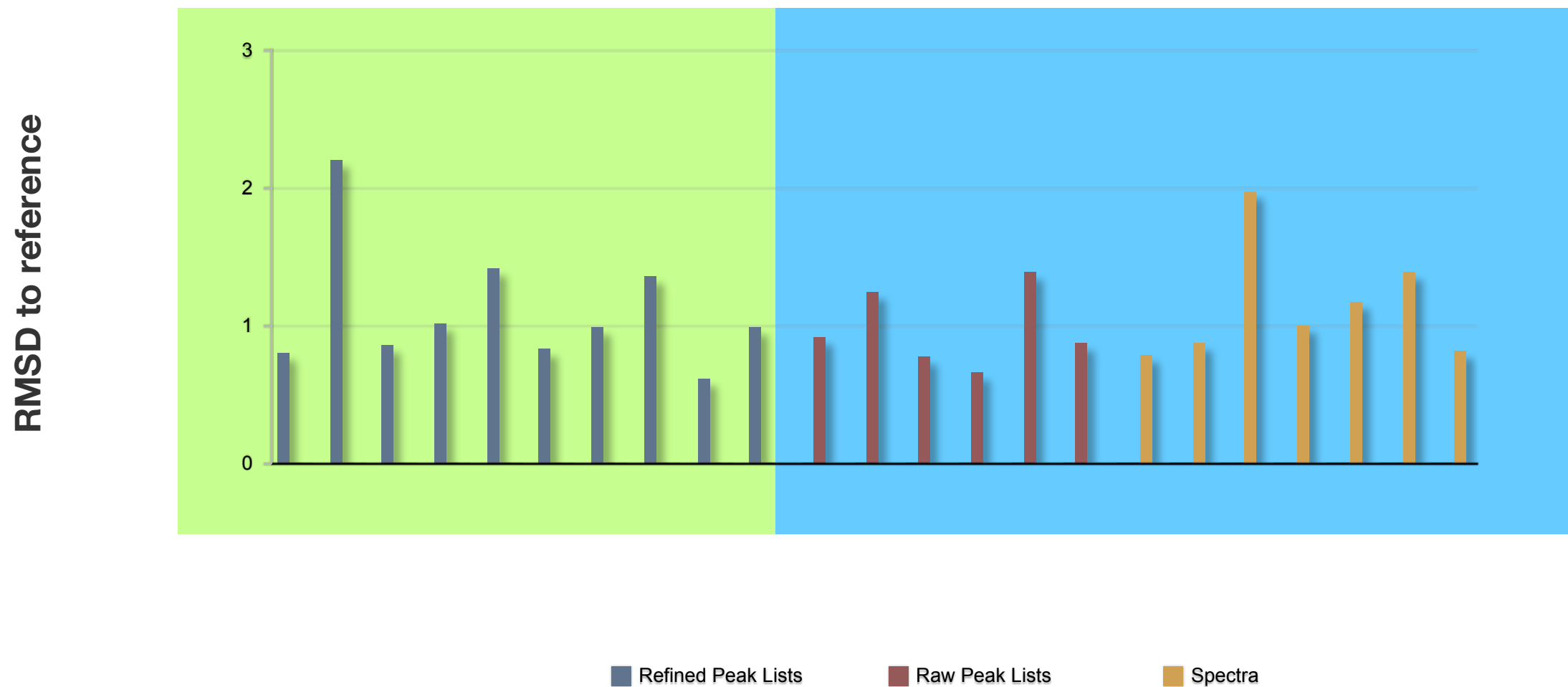
“It can be concluded that **NOESY-based methods** delivered **more consistent and robust performances** than CS-based methods, yielding structures **on average closer to the reference**. NOESY-filtering as in CS-DP-Rosetta could recover some but not all of the consistency and reliability of the restraint-driven methods (discussed later). **Notably, the CS-methods** (regardless of whether augmented with NOESY information) are **computationally much more demanding** than NOESY-based methods.”

From Structure 2012, 20(2): 227-236.

# CASD I&2-NMR: UNIO'12 performance

CASD-NMR 1

CASD-NMR 2



Perfect world

Closer to real world

$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} \frac{\hbar^2 \gamma_1 \gamma_2}{3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt} \sigma = -i [\mathcal{H}, \sigma]$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} \frac{\hbar^2 \gamma_1 \gamma_2}{3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

# Outline - Part 2



**UNIO**

**UNIO'13 - Integration and modeling**



$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

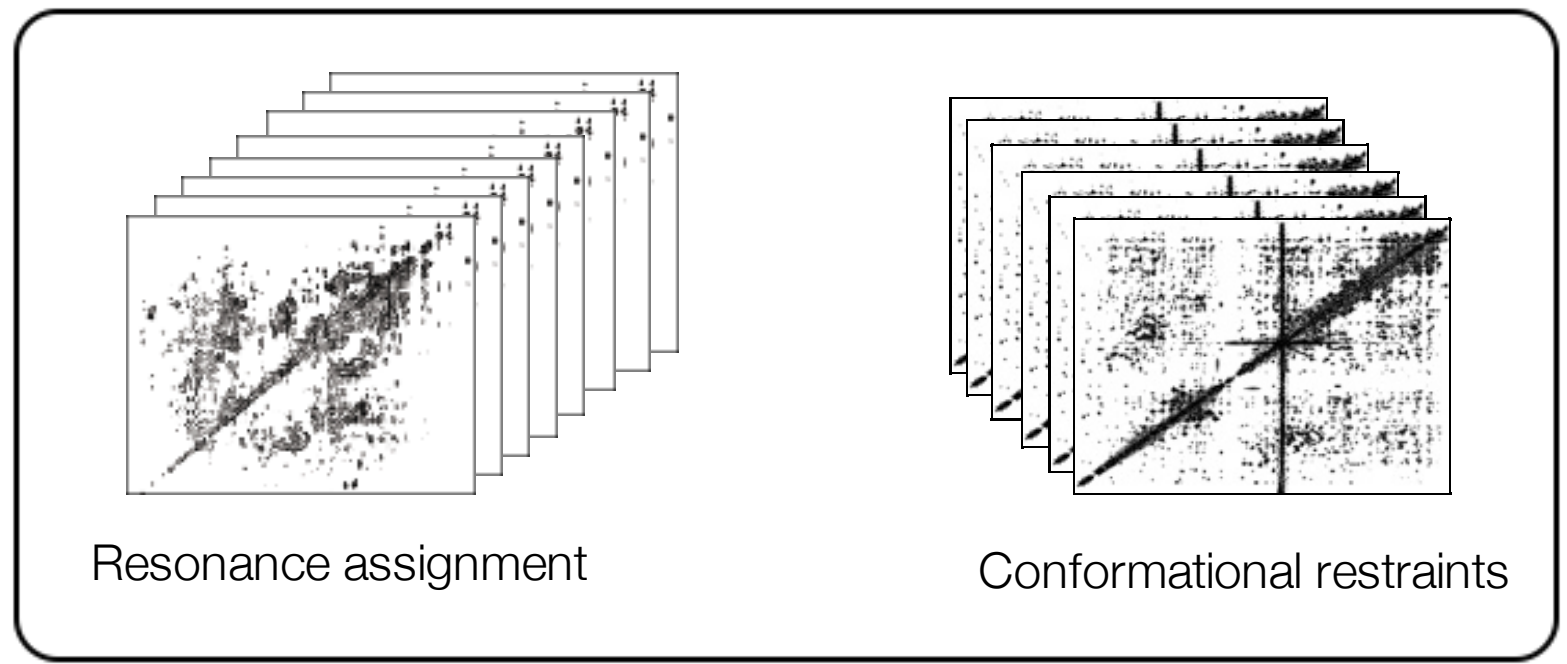
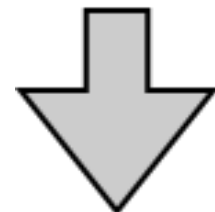
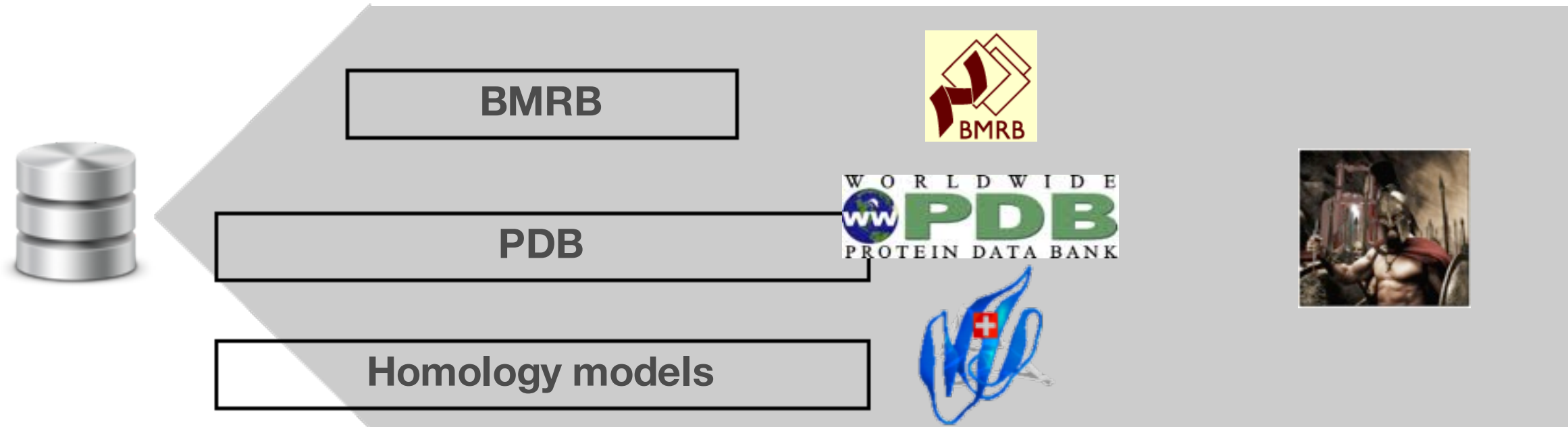
$$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda} \frac{\hbar^2 \gamma_1 \gamma_2}{\pi r^3} (1 - 3 \cos^2 \theta) (3 I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt} \sigma = -i [\mathcal{H}, \sigma]$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda} \frac{\hbar^2 \gamma_1 \gamma_2}{\pi r^3} (1 - 3 \cos^2 \theta) (3 I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

# How to use data base information?

## Correctness?



Resonance assignment

Conformational restraints

$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$

$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} \frac{\hbar^2 \gamma_1 \gamma_2}{3 \cos^2 \theta} (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$

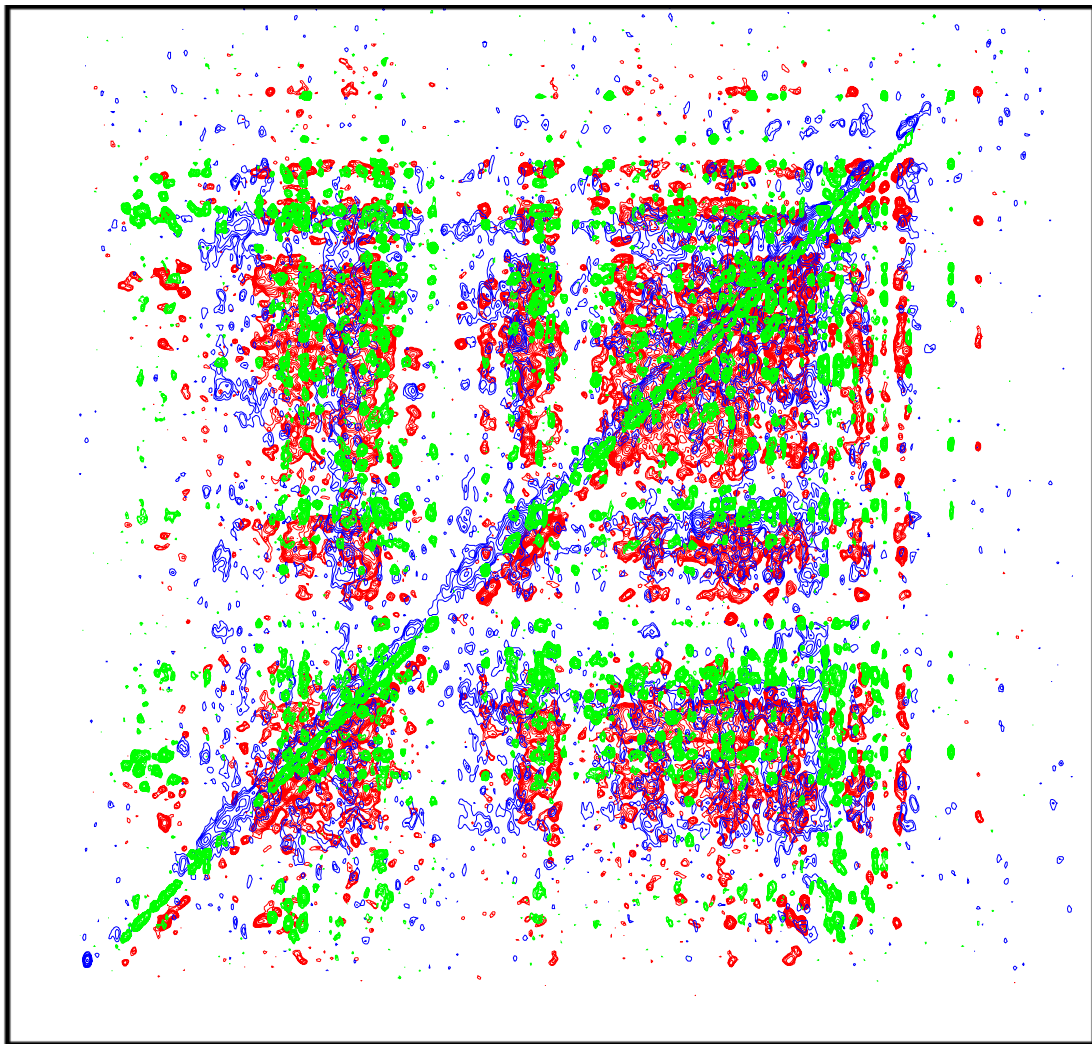
$\frac{d\sigma}{d\Omega} = -i[\mathcal{H}, \sigma]$

$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} \frac{\hbar^2 \gamma_1 \gamma_2}{3 \cos^2 \theta} (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$

$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} \frac{\hbar^2 \gamma_1 \gamma_2}{3 \cos^2 \theta} (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$

$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} \frac{\hbar^2 \gamma_1 \gamma_2}{3 \cos^2 \theta} (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$

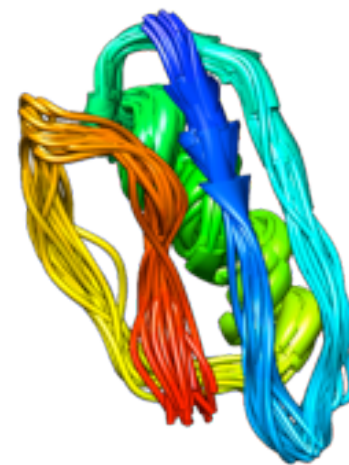
*Data base information can be dangerous!*



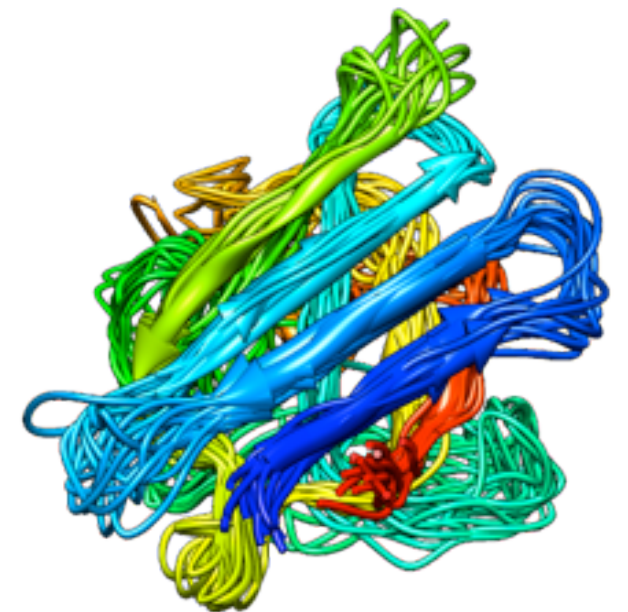
Ubiquitin

GB1

SOD



GB1 from ubiquitin spectra  
rmsd to reference: 1.7 Å



SOD from GB1 & ubiquitin spectra  
rmsd to reference: 2.8 Å

*Prerequisite for the application of data base knowledge:*

*Correct chemical shift referencing*



**UNIO-MATCH**

- LACS: Wang et al. (2005) J. Biomol. NMR 32.
- CheckShift: Simon et al. (2007) J. Biomol. NMR 39.
- PANAV: Wang et al. (2010) J. Biomol. NMR 47.
- VASCO: Rieping et al. (2010) Proteins 78.
- RefDB: Zhang et al. (2003) J Biomol NMR 25.

### Potential weaknesses of current procedures:

- ◆ Only applicable to a subset of backbone atoms.
- ◆ Strong assumptions on correct referencing of selected backbone shifts.
- ◆ Structure needed as input.

### Not Found

The requested URL ██████████ was not found on this server.

Additionally, a 404 Not Found error was encountered while trying to use an ErrorDocument to handle the request.

$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$

# Prerequisite for the application of data base knowledge:

$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} \frac{\hbar^2 \gamma_1 \gamma_2}{\dots}$

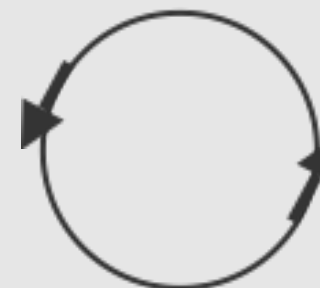
## Correct chemical shift referencing

$\frac{d\sigma}{dt} = -\rho(\sigma)$   
 $(1-3\cos^2\theta)(3I_{1z}I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$

- Data base of average values for secondary chemical shifts of atoms in protein (coil, strand, helix).

(e.g. Wang & Jardetzky Protein Sci. 2002)

- For each residue, determine the secondary structure elements using the joint probability over CA, CB, CO, N, HN, HA shift values.
- Determine **individually** for each backbone atom the offset between input chemical shift value and derived secondary distribution function.



- ★ **Crucial merit 1: chemical shifts do not need to be assigned**, i.e., procedure applicable at the outset of a structure determination process.
- ★ **Crucial merit 2:** Chemical shift correction and standard deviations is given.
- ★ **Now:** For backbone resonance assignment a safe comparison to chemical shift statistics can be done.

$$\omega_1, \omega_2) = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

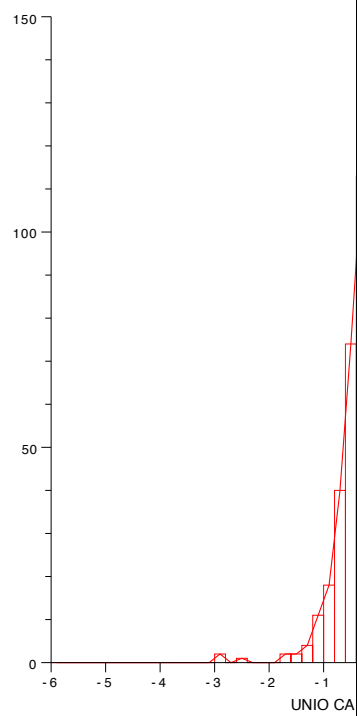
$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_N \mu_N}{4\pi r^3} (1 - 3\cos^2\theta)$$

$$\frac{d\sigma}{d\Omega} = f(\theta, \phi)$$

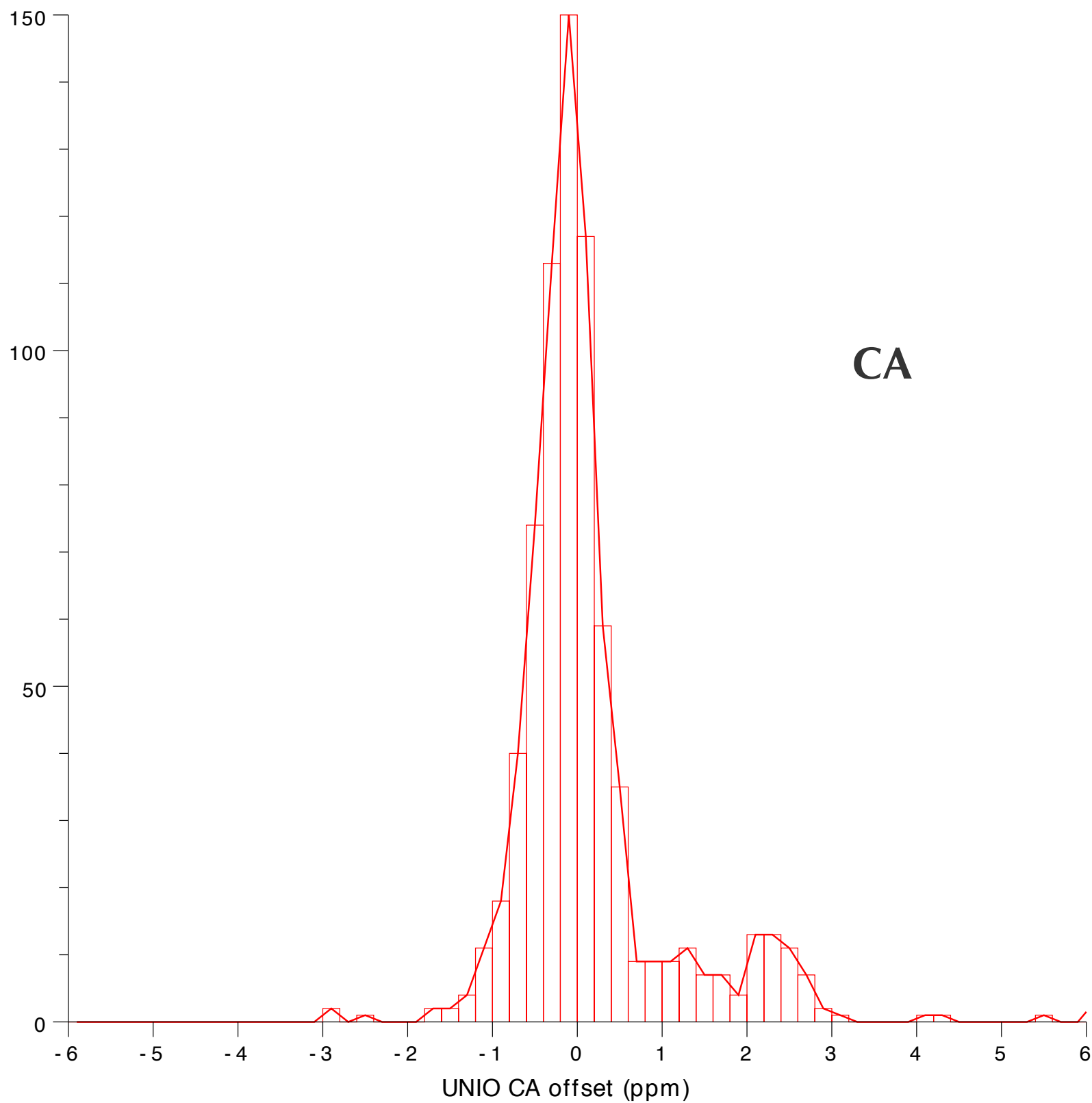
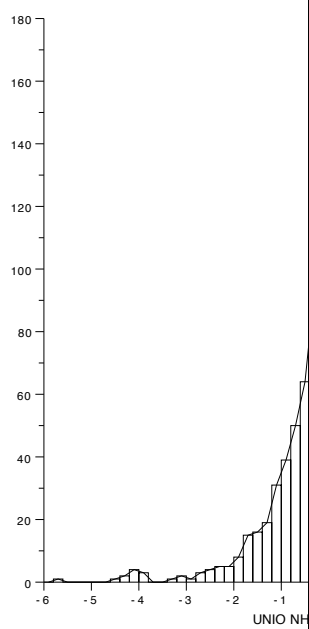
$$I(\omega_1, \omega_2) = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_N \mu_N}{4\pi r^3} (1 - 3\cos^2\theta) (3I_{1z}I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

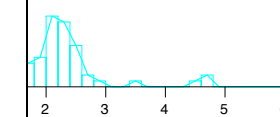
Distribution of CA



Distribution of NH



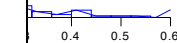
offset in the BMRB



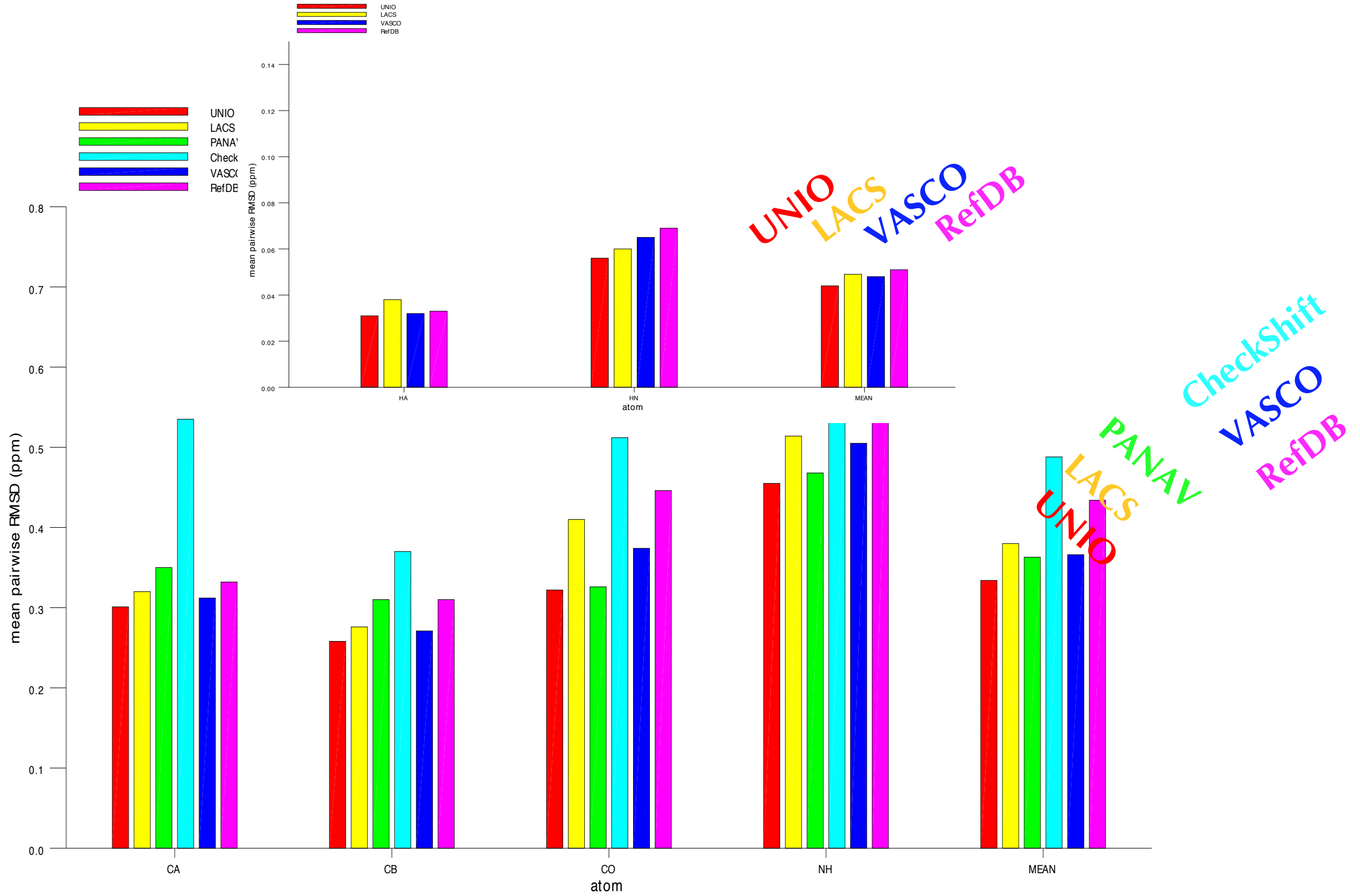
CO

offset in the BMRB

HA

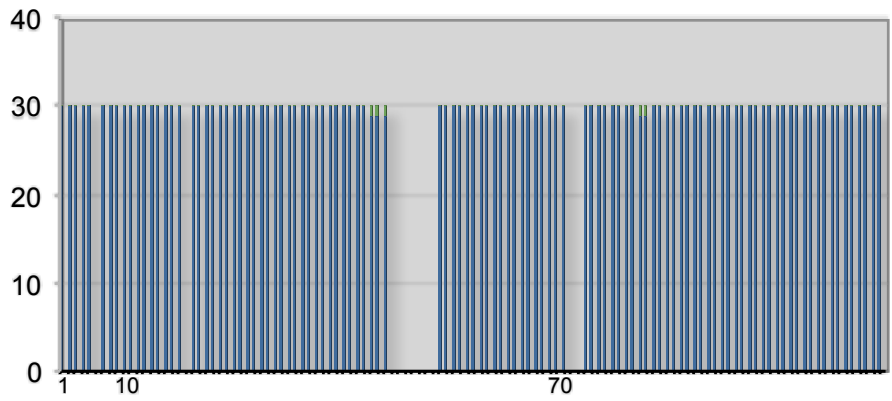
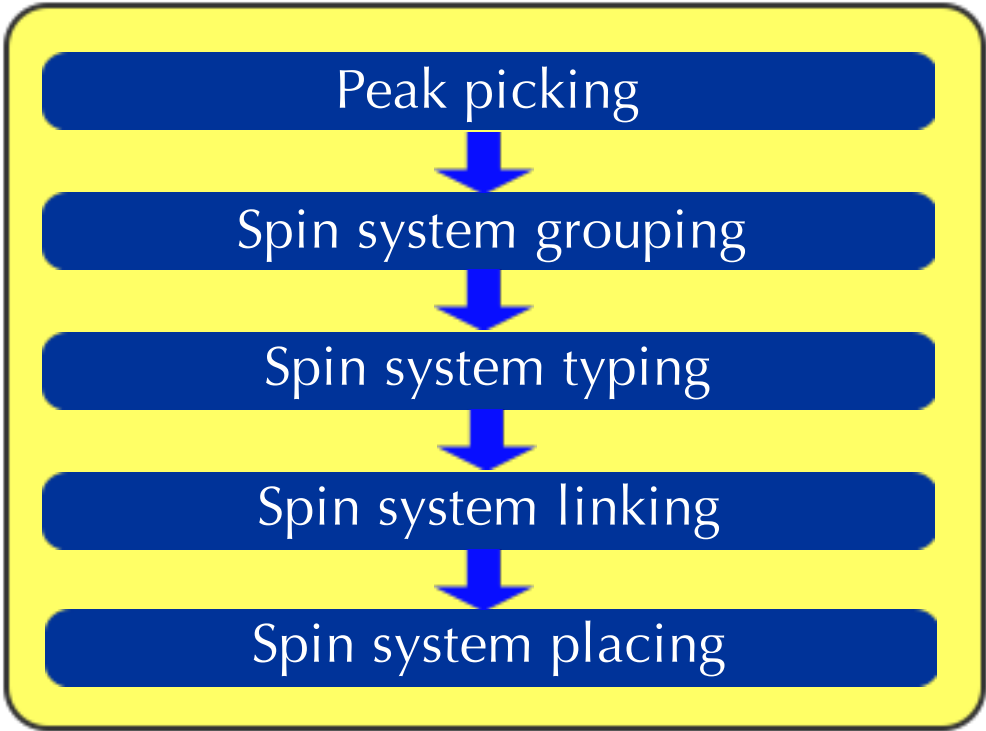


# UNIO RefBMRB - Comparison to other programs

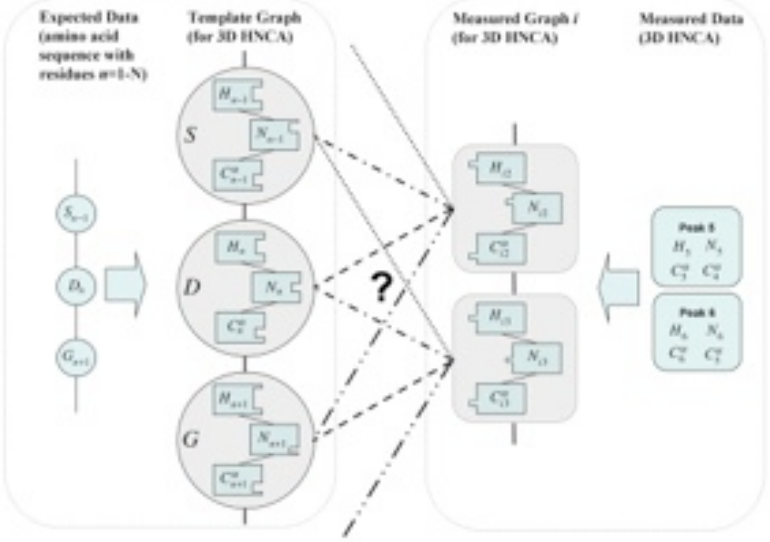


# I. Integration & modeling: Backbone resonance assignment

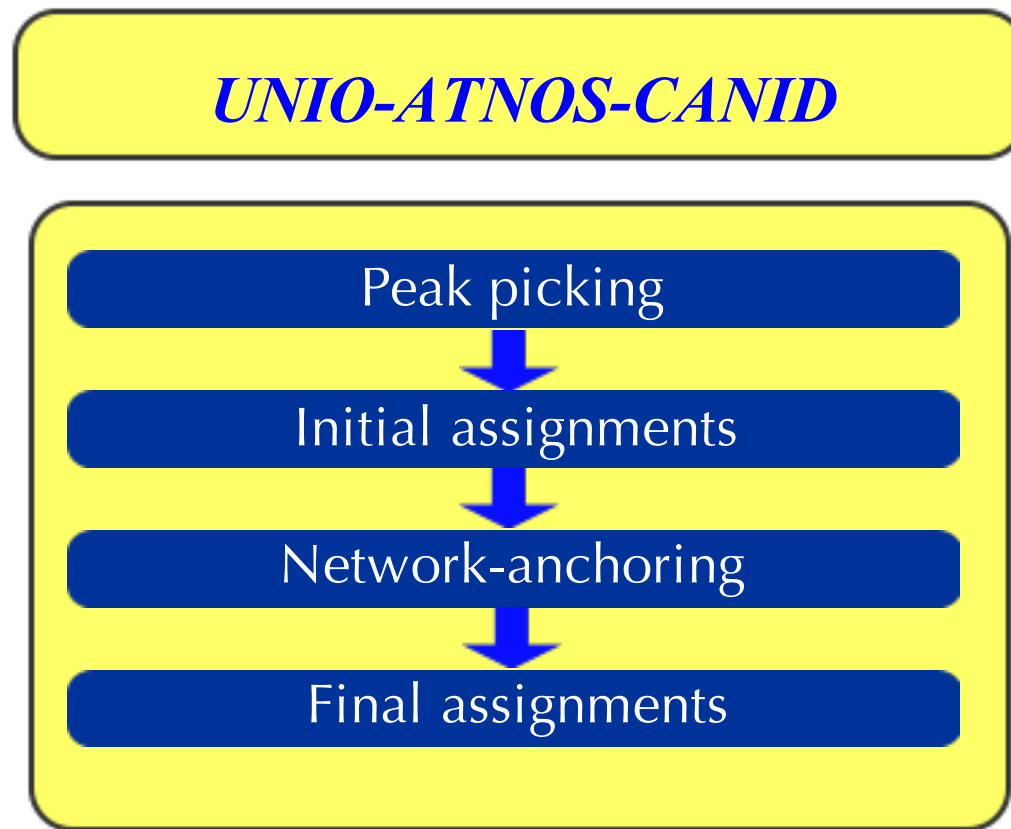
**UNIO-MATCH**



al assignments      ■ Missing assignments



## 2. Integration & modeling: NOE assignment



- ★ **Principal idea 1:** Retain structure-based assignments together with other assignment possibilities in the first round of NOE assignment.
- ★ **Principal idea 2:** Either structure-based assignments are in agreement with experimental data; if not, algorithm will converge to the 'real' structure. Retained structure-based assignments were only an initial disturbance of the procedure.

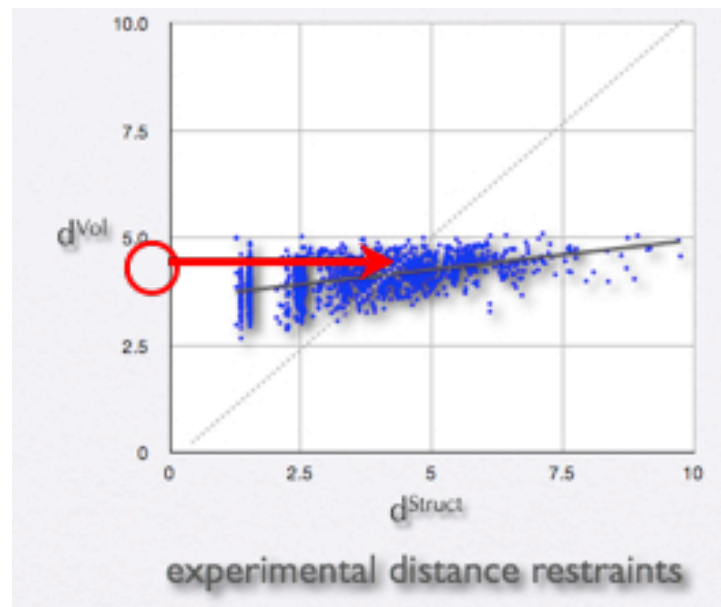
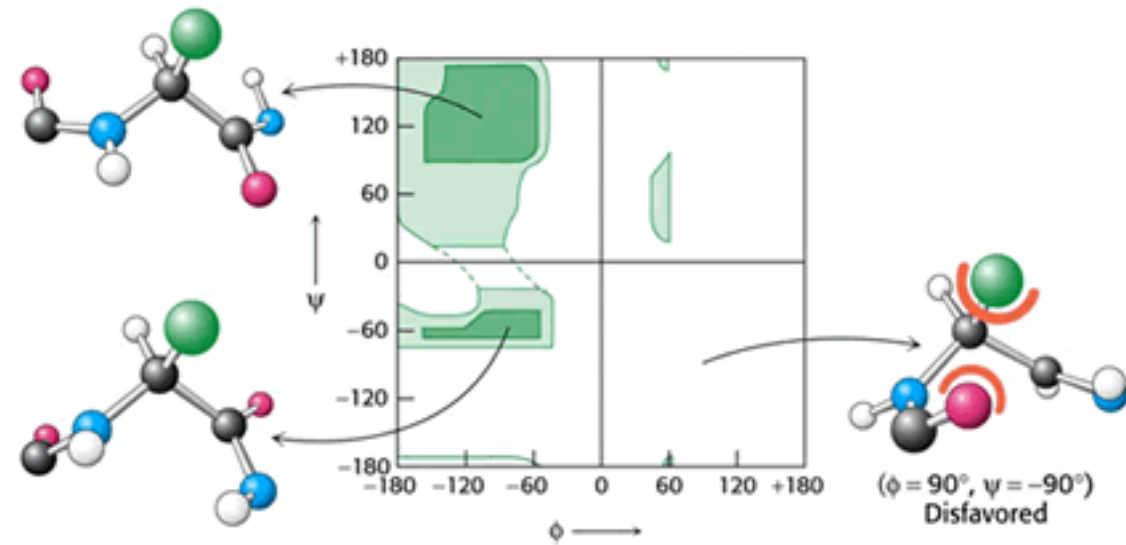


# 3. Integration & modeling: Distance estimations by exhaustive grid search

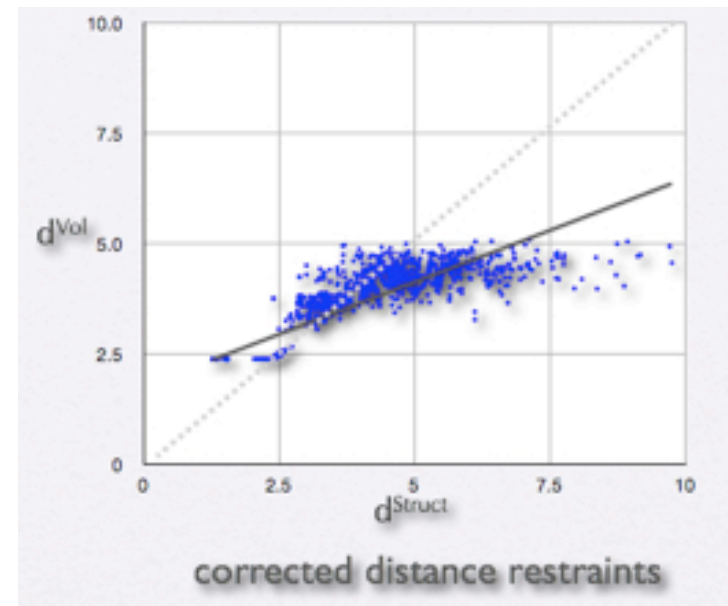
Correct upper bounds for distance restraints.

## Distance estimation in proteins

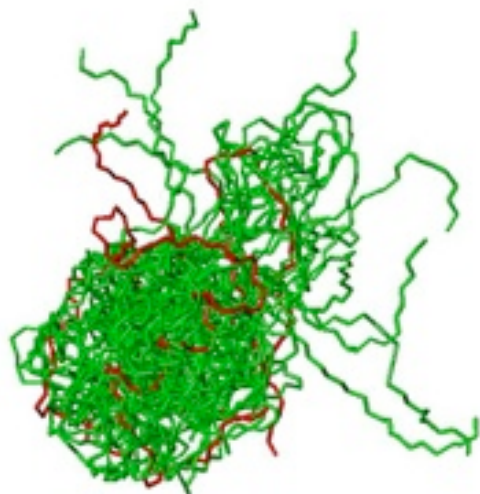
- Exhaustive conformational sampling.
- Incorporation of a **priori knowledge**.



Without



With



$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} \frac{\hbar^2 \gamma_1 \gamma_2}{3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt} \sigma = -i [\mathcal{H}, \sigma]$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu}{\lambda \pi r^3} \frac{\hbar^2 \gamma_1 \gamma_2}{3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt} \sigma = -i [\mathcal{H}, \sigma]$$

# Outline - Part 3



**UNIO**

**UNIO'13 - Integration and modeling**

**Nanomachineries**

# Solid state NMR structure determination

## Micro-crystalline proteins

GBI (2005, 2008, 2009) Ubiquitin (2008)



CrH (2008)



Kaliootoxin (2005) SH3 domain (2002)



MMP-12 (2009)



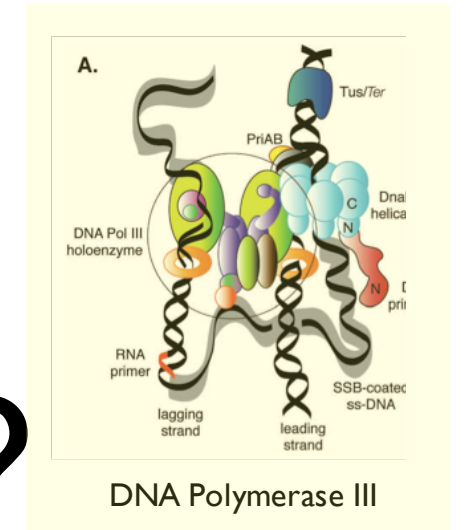
## Next targets



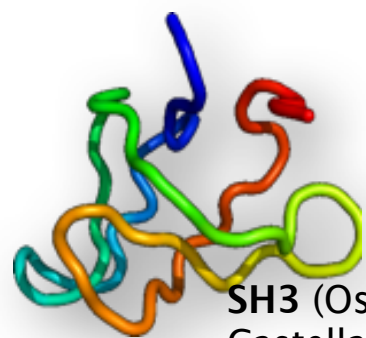
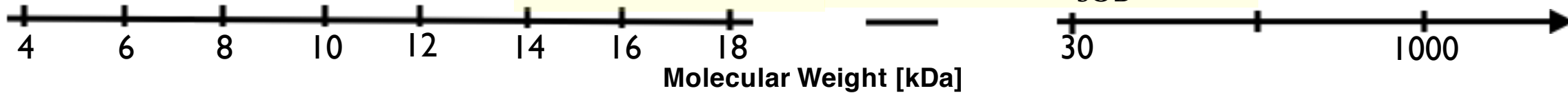
$\alpha\beta$  crystalline (2010)



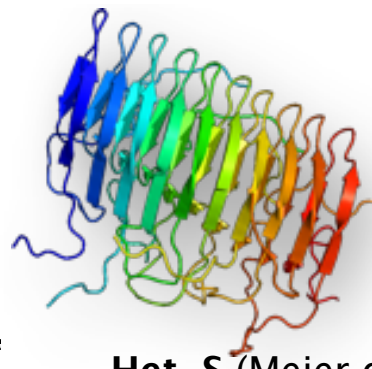
SOD



?



SH3 (Oschkinat group),  
Castellani et al.  
Nature 2002

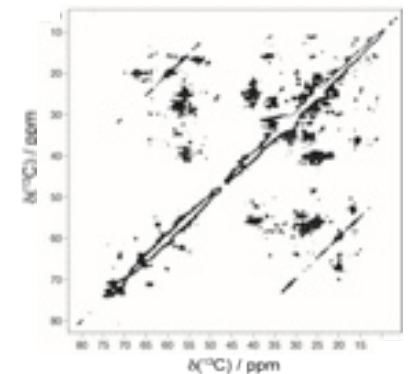


Het-S (Meier group),  
Wasmer et al.  
Science 2008



Type-III Secretion system  
(Lange group) Nature 2012

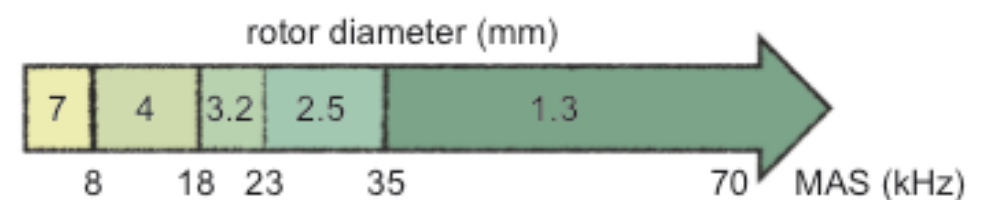
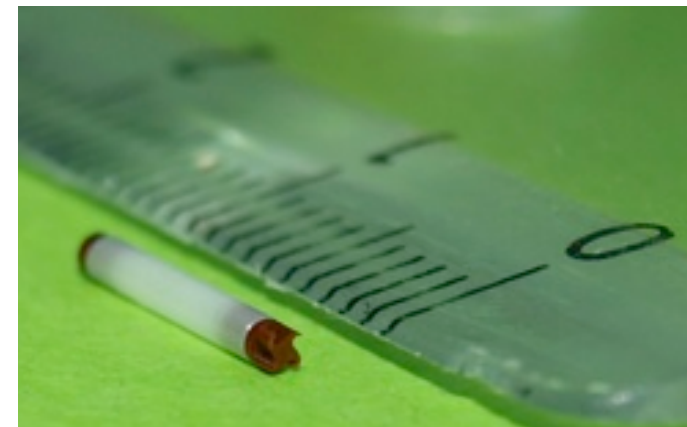
Virus nucleocapsid:



GHz spectra of GigaDalton molecules

# Proton detection, high magnetic field & ultra-fast MAS

- ◆ All results were recorded using a 1 GHz instrument and 60 kHz MAS at CRMN, ENS-Lyon
- ◆ 100% back-protonation was used in all experiments



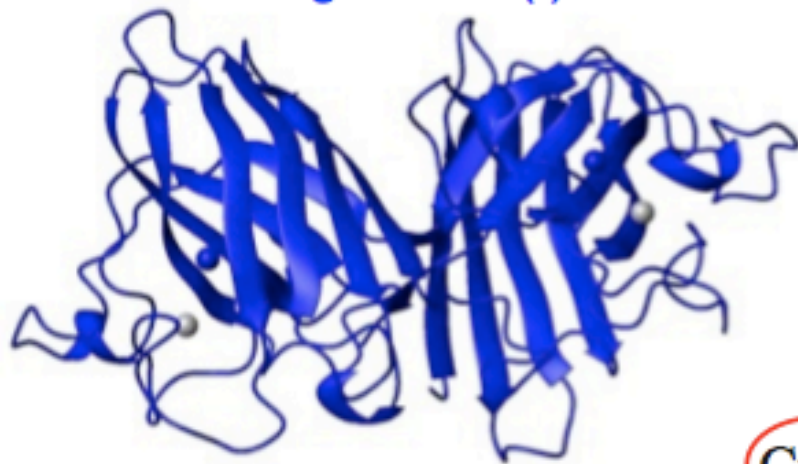
$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$(\omega_1, \omega_2) = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

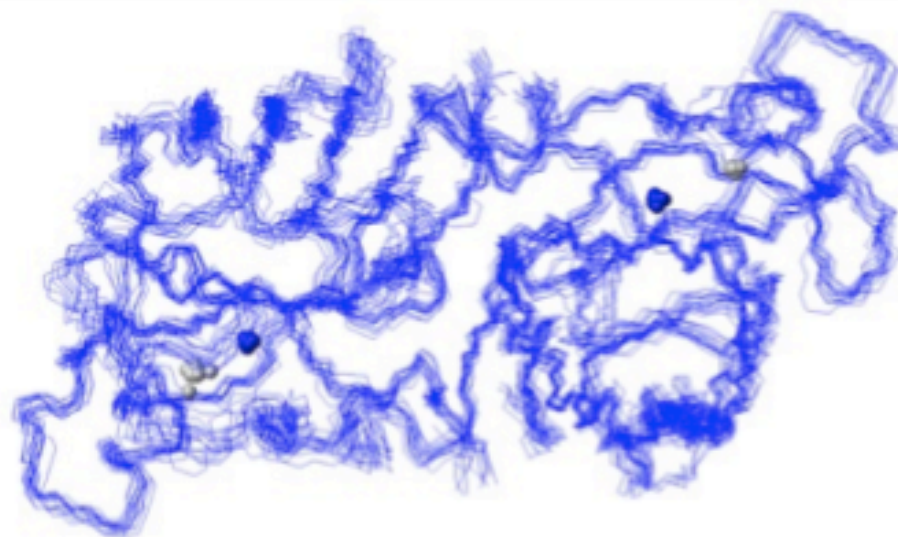
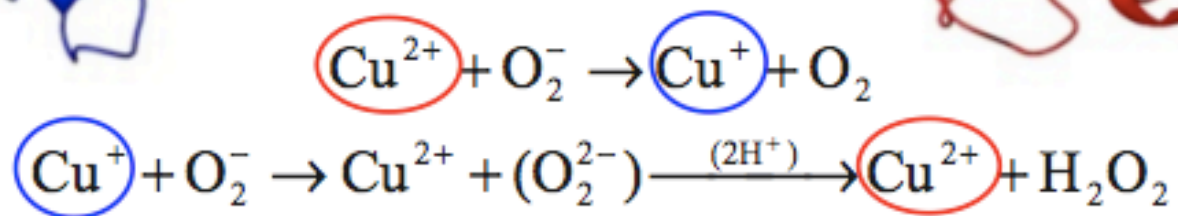
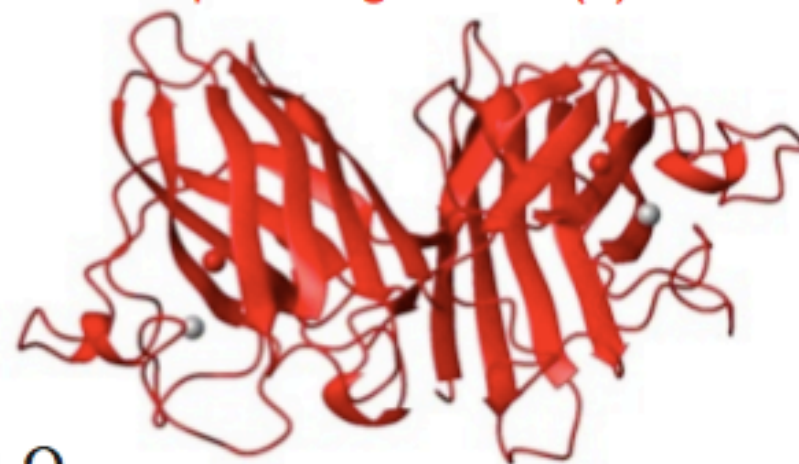
# Model system: Human superoxide dismutase (32kDa dimer)

## Human CU(II), Zn(II)-superoxide dismutase (32kDa)

diamagnetic Cu(I)



paramagnetic Cu(II)



Banci et al., *Eur. J. Biochem.* 2002

Sample: non-diffracting nanocrystals

$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \gamma^2 \gamma'}{4\pi r^3} (1-3\cos^2\theta)(3I_{1z}I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt}\sigma = -i[\mathcal{H}, \sigma]$$

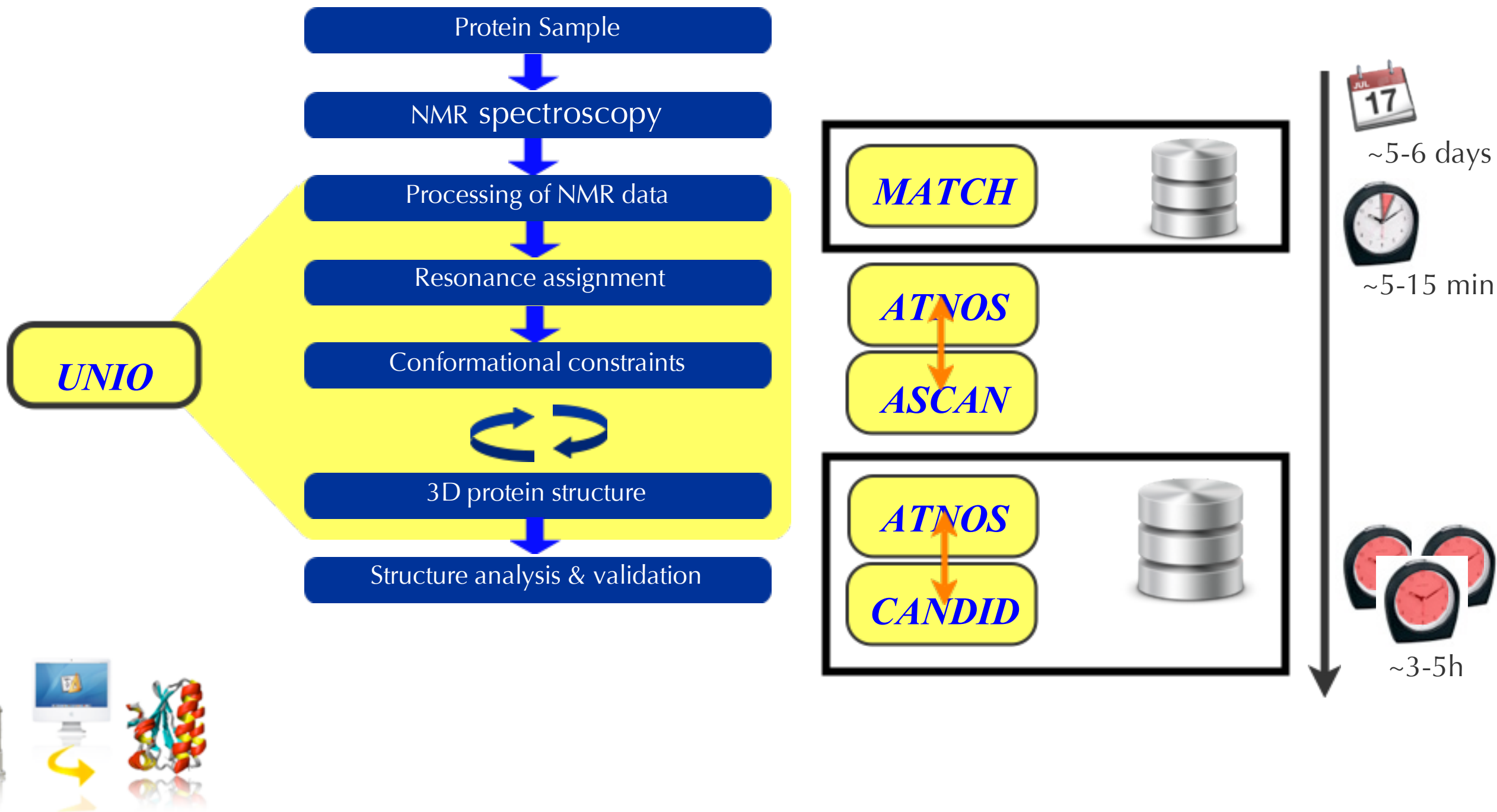
UNIO '13

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \gamma^2 \gamma'}{4\pi r^3} (1-3\cos^2\theta)(3I_{1z}I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt}\sigma = -i[\mathcal{H}, \sigma]$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \gamma^2 \gamma'}{4\pi r^3} (1-3\cos^2\theta)(3I_{1z}I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

# NMR data analysis using comparative modeling



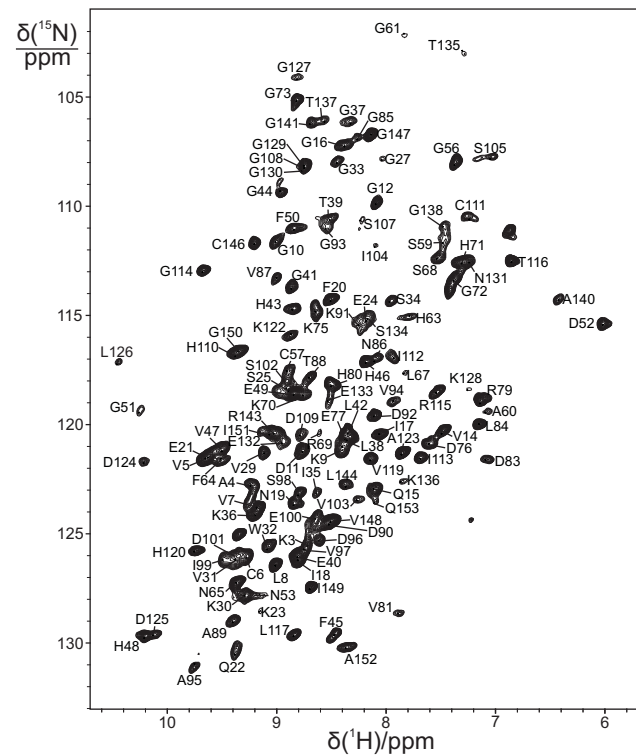
$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$

# Resonance assignment & distance restraints using proton detection

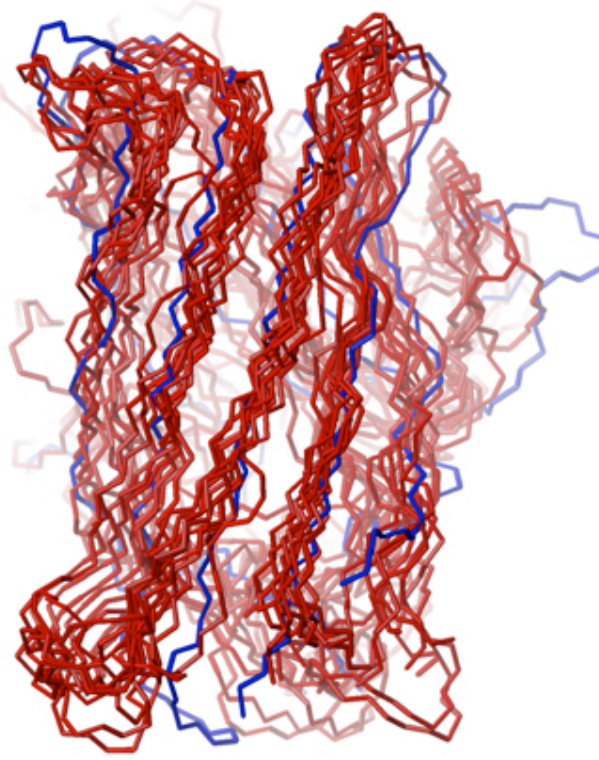
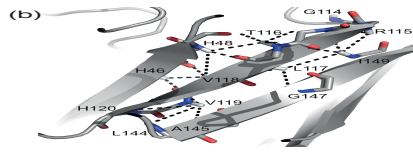
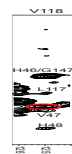
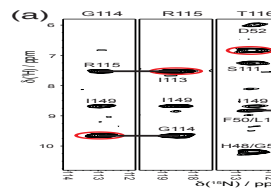
$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3 I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$

$\mathcal{H}_{\sigma} = \frac{d\sigma}{d\Omega} \frac{1}{r^2} \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\}$

$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3 I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$



+



- ◆ Using UNIO-MATCH and UNIO-ATNOS/CANDID: 192 1H-1H restraints
- ◆ RMSD to mean: 1.64 Å
- ◆ RMSD to X-ray: 2.34 Å

$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$

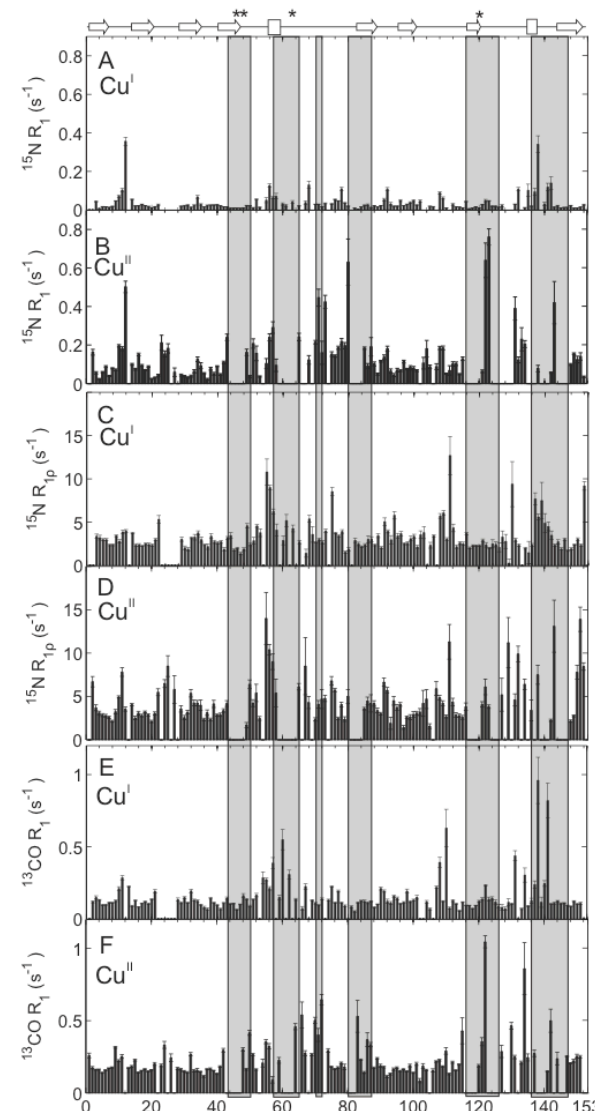
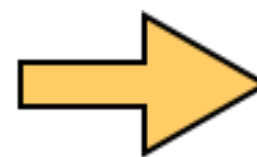
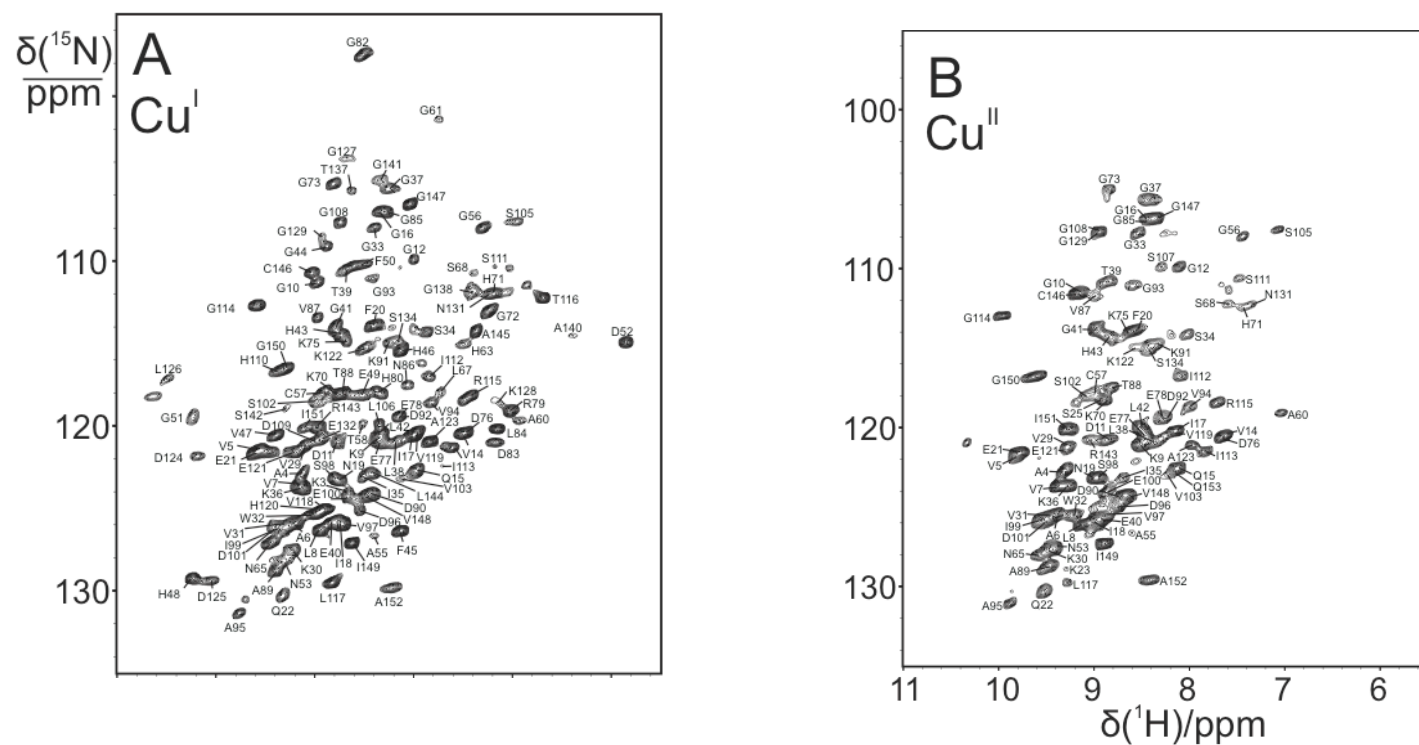
$\mathcal{H}_D = \frac{1}{2} \frac{\mu_N \hbar^2 \gamma_1 \gamma_2}{\lambda \pi r^3} (1 - 3 \cos^2 \theta) (3 I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$

$\frac{d\sigma}{dt} = -i[\mathcal{H}, \sigma]$

$\mathcal{H}_D = \frac{1}{2} \frac{\mu_N \hbar^2 \gamma_1 \gamma_2}{\lambda \pi r^3} (1 - 3 \cos^2 \theta) (3 I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$

# Site-specific backbone dynamics

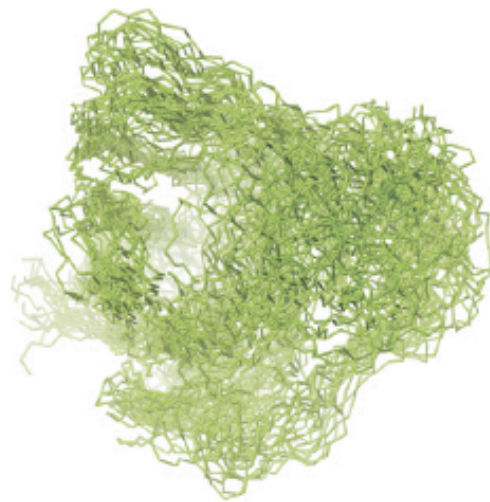
- ◆ H-H spin-diffusion (RDFR) distance restraints -> short-range distance restraints
- ◆ Paramagnetic relaxation enhancements (PRE) -> long-range distance restraints
- ◆ Gaussian axial fluctuation (GAF) analysis -> order parameters and timescales



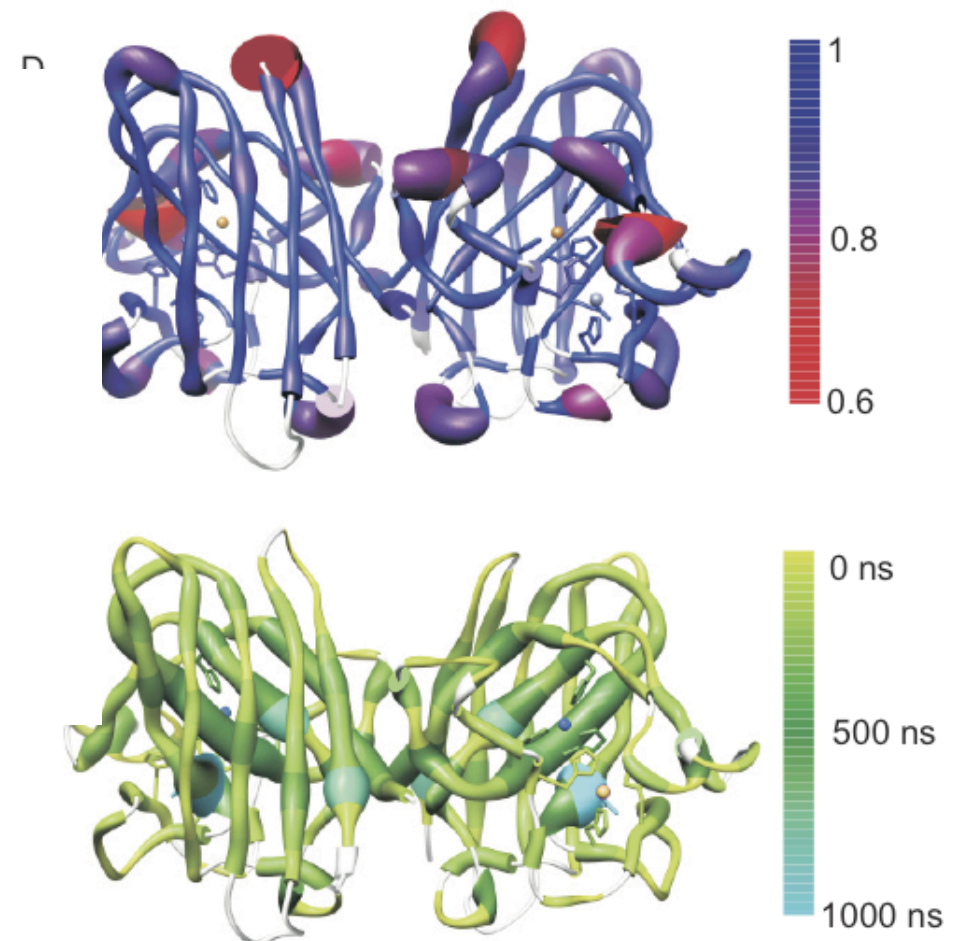
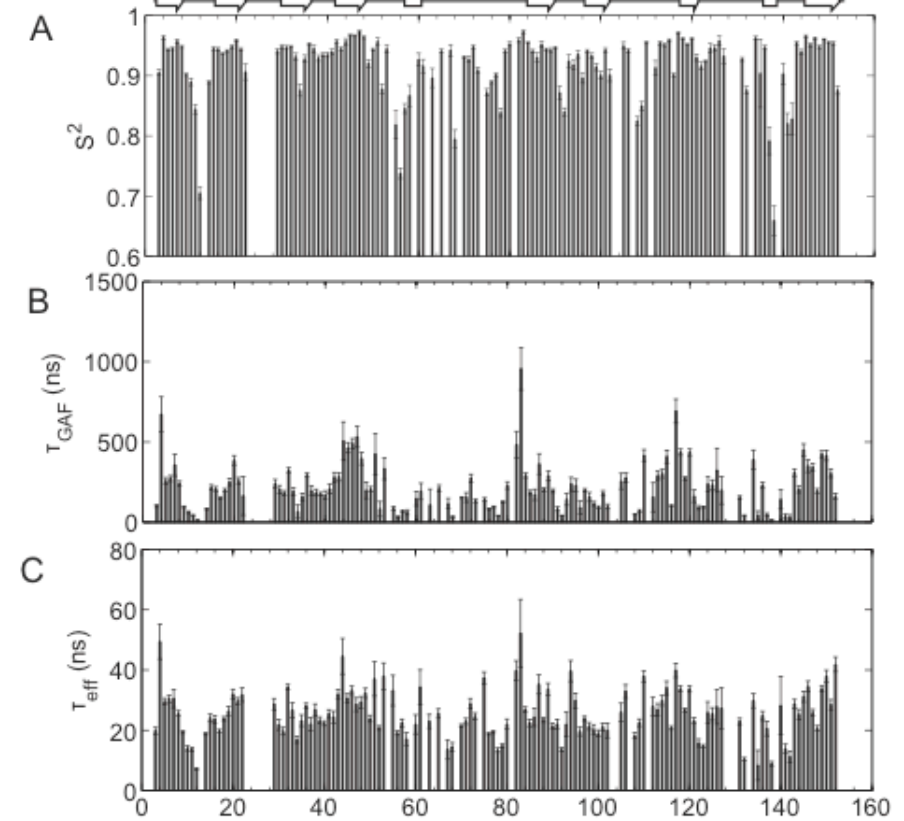
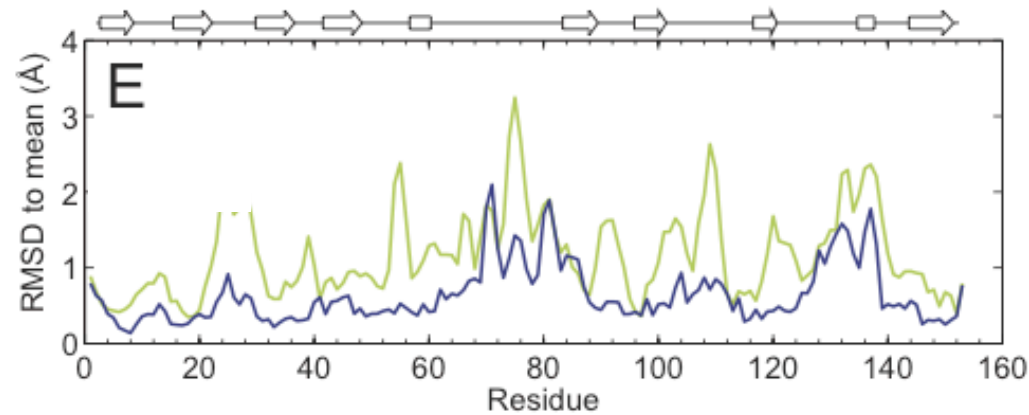
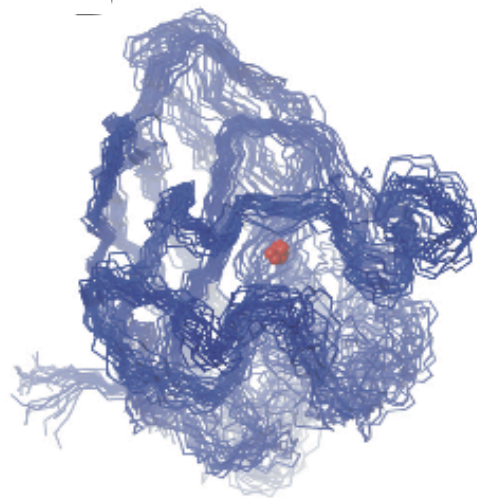
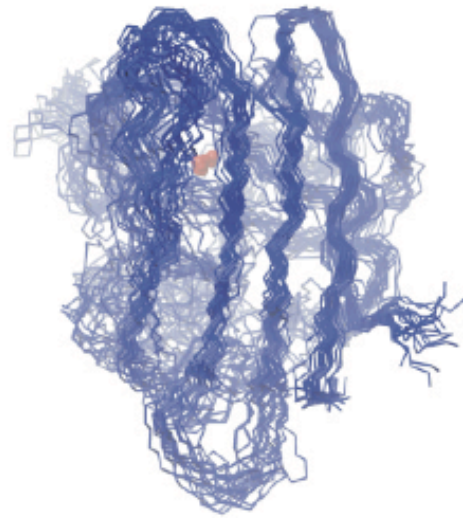


# Refined SOD structure and dynamics using PRE

w/o PRE



w PRE



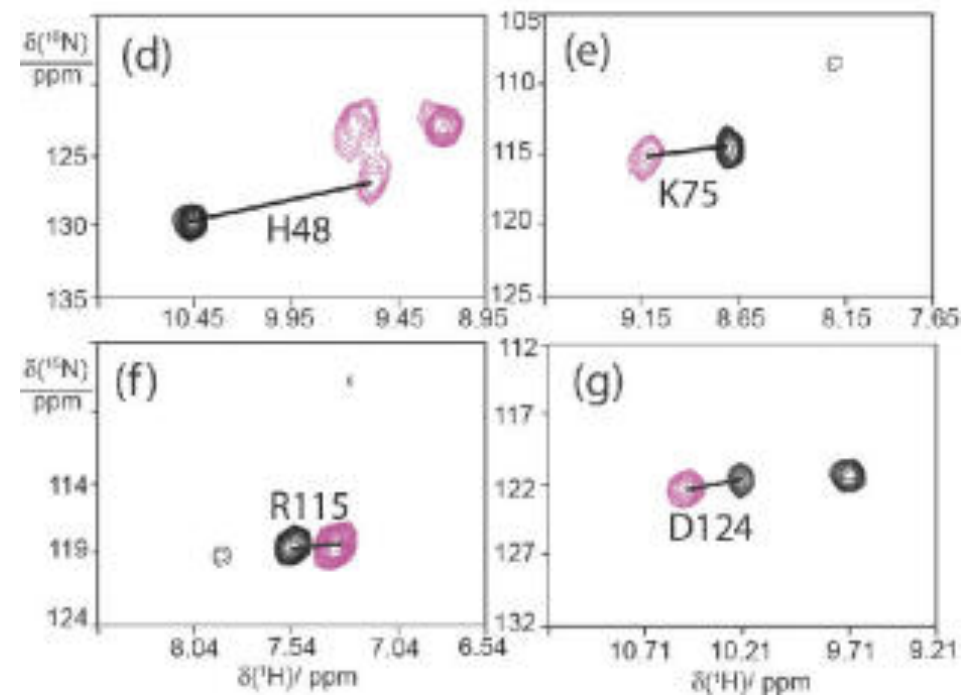
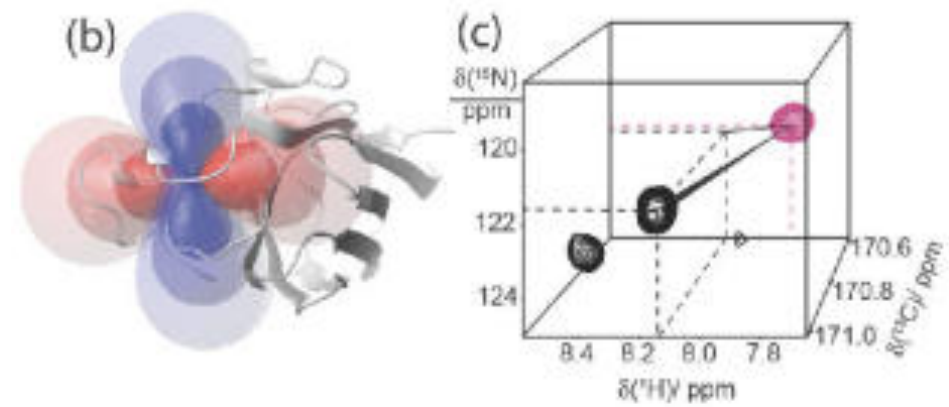
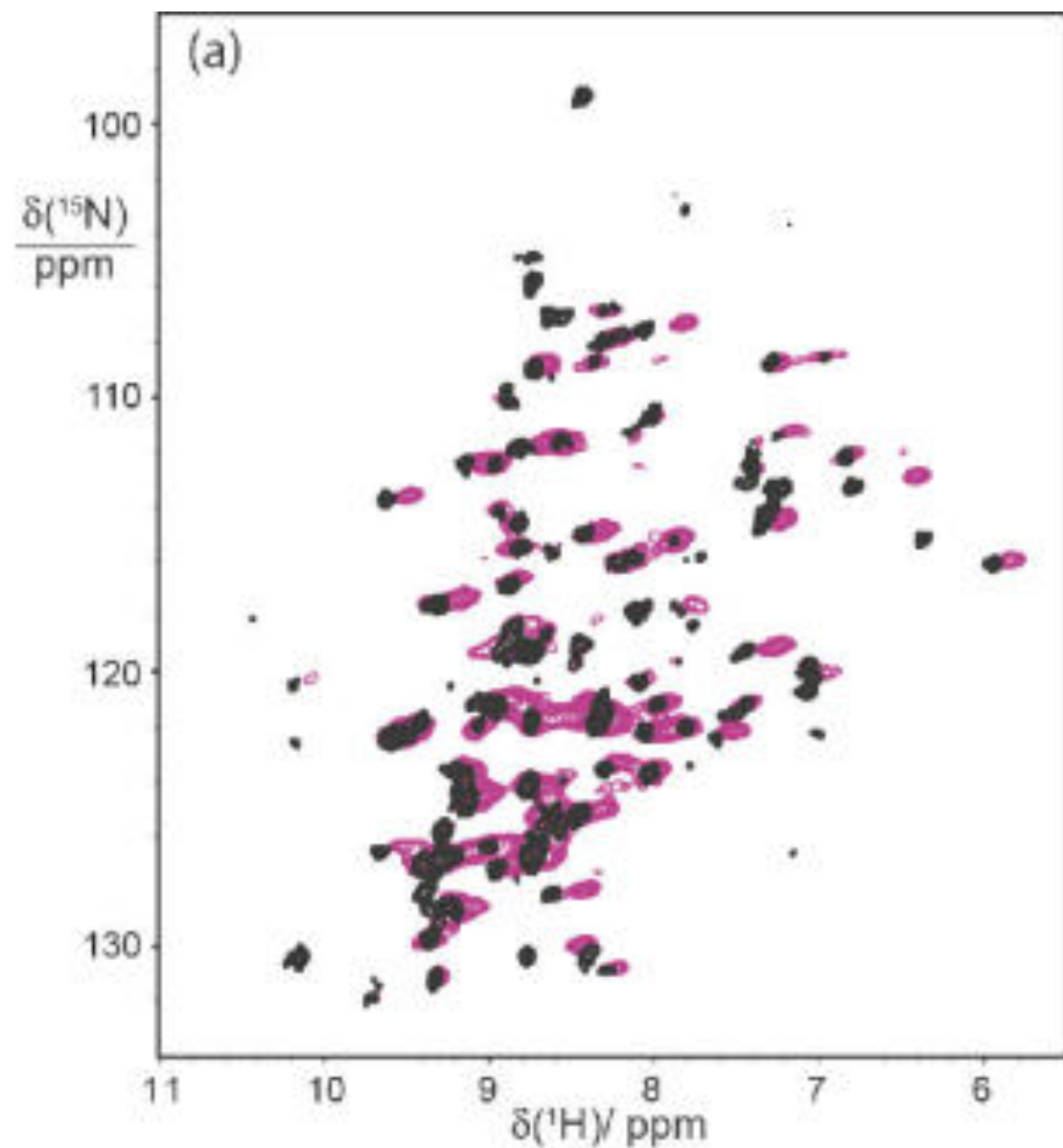
$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3 I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt} \sigma = -i [\mathcal{H}, \sigma]$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3 I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

# Pseudo contact shifts in SOD



$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$

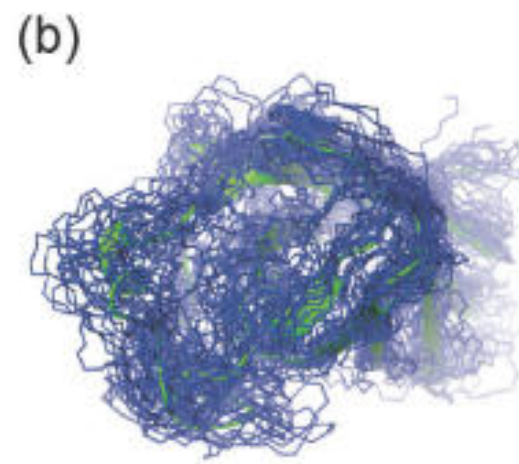
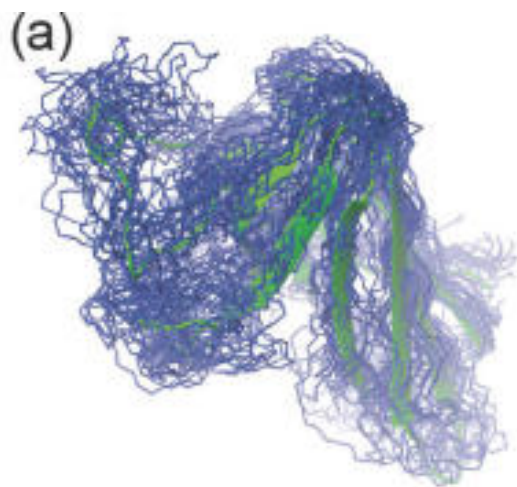
$\mathcal{H}_D = \frac{1}{2} \frac{\mu_N^2 \gamma_1 \gamma_2}{\lambda \pi r^3} (1 - 3 \cos^2 \theta) (3 I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$

$\frac{d\sigma}{dt} = -i[\mathcal{H}, \sigma]$

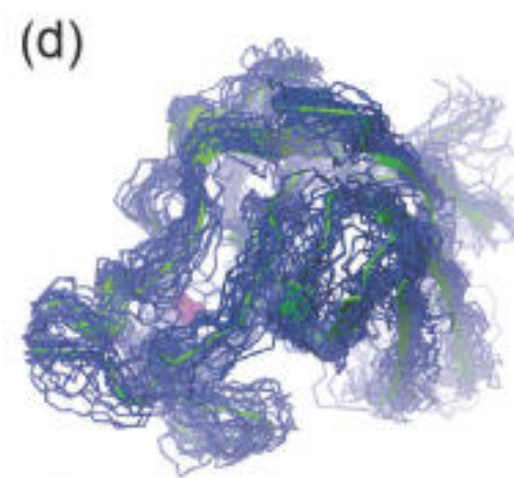
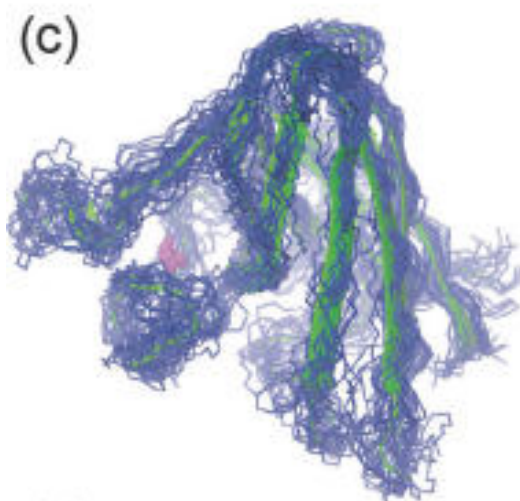
$\mathcal{H}_D = \frac{1}{2} \frac{\mu_N^2 \gamma_1 \gamma_2}{\lambda \pi r^3} (1 - 3 \cos^2 \theta) (3 I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$

# SOD structure using diamagnetic restraints, PRE and PCS

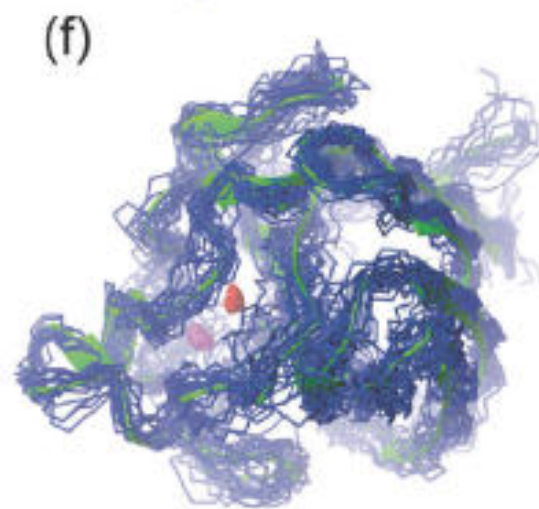
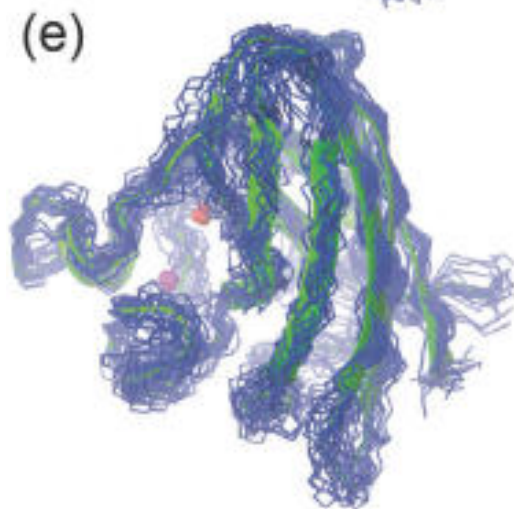
1H-1H only



1H-1H + PCS



1H-1H + PCS + PRE



$$\omega_1, \omega_2 = \iint s(t_1, t_2) \exp\{-i(\omega_1 t_1 + \omega_2 t_2)\} dt_1 dt_2$$

$$\mathcal{H}_D = \frac{1}{2} \frac{\mu_0 \hbar^2 \gamma_1 \gamma_2}{4\pi r^3} (1 - 3 \cos^2 \theta) (3I_{1z} I_{2z} - \vec{I}_1 \cdot \vec{I}_2)$$

$$\frac{d}{dt} \sigma = -i [\mathcal{H}, \sigma]$$

# Acknowledgment

## UNIO and J-UNIO protocol

## Solid State NMR

Paul Guerry



Michael Knight

Guido Pintacuda

CERM, Florence, Italy

Paul Guerry

Jozef Lewandowski

Benedicte Elena

Moreno Lelli

Anne Lesage

Lyndon Emsley

